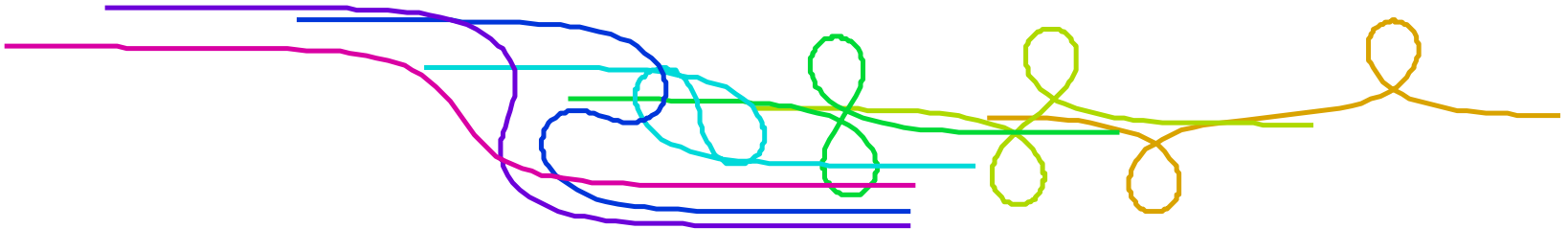


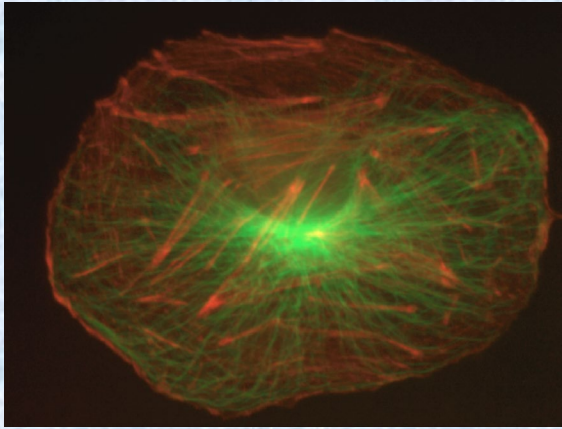
Chem 163

10 Nov. 2022
Adam E. Cohen

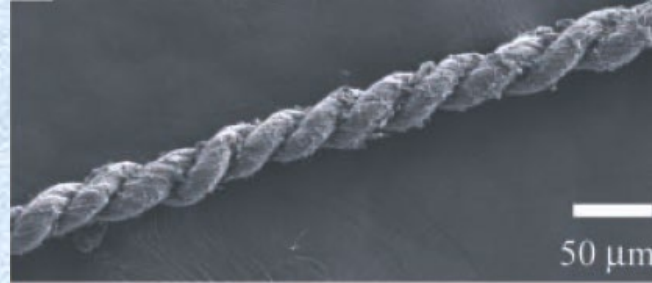


Mechanics of rods

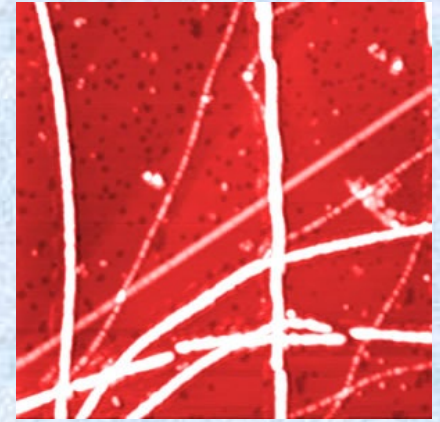
Fibrils and Us



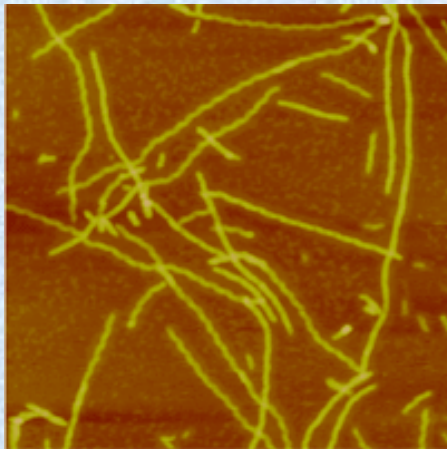
actin & tubulin



rope of carbon nanotubes



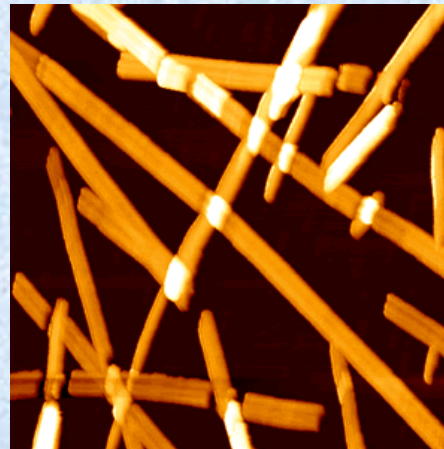
j-aggregates



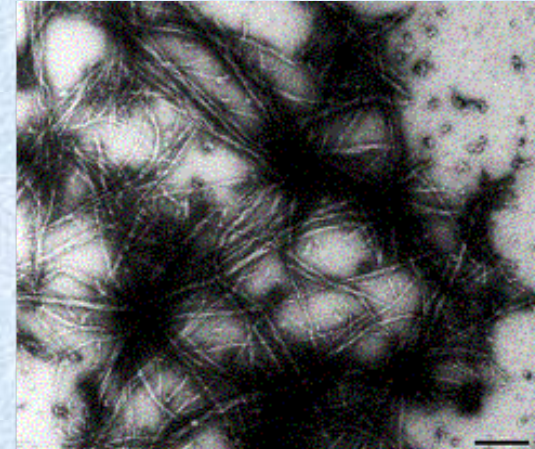
prions



sickle-cells

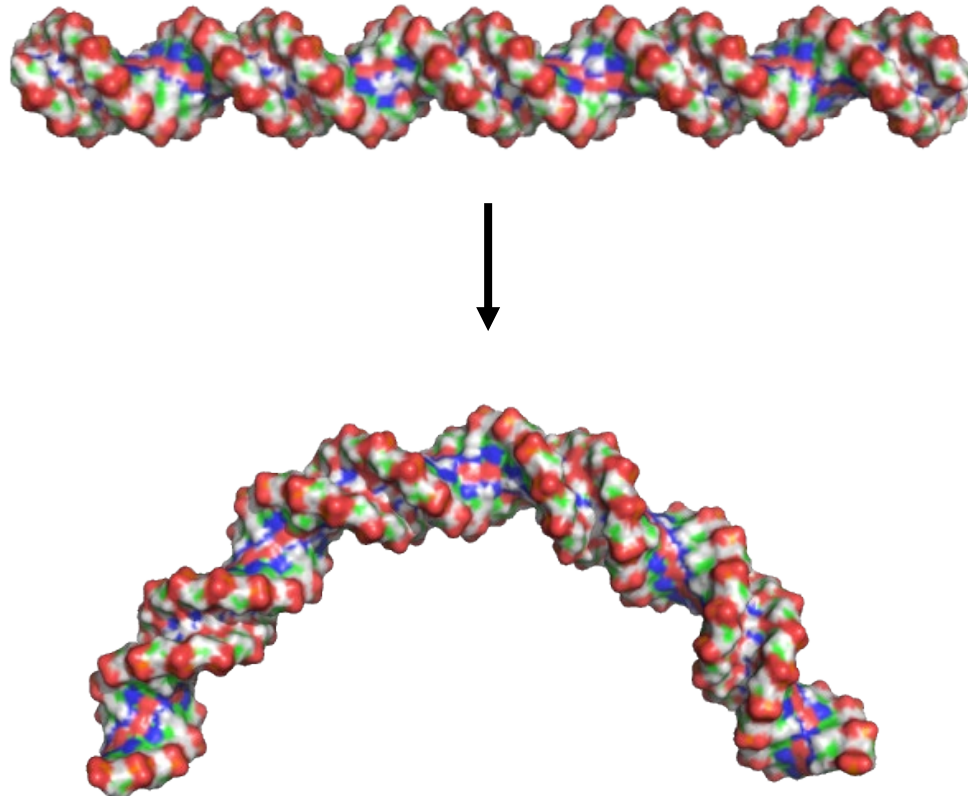


amyloids



cataracts

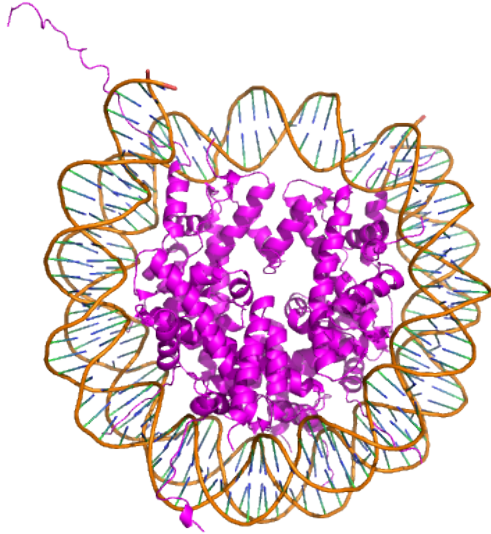
DNA Bending



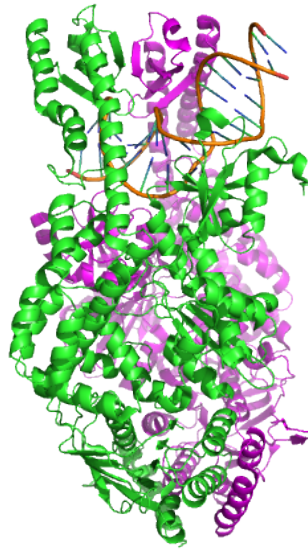
Nucleic Acid Builder (NAB)

Macke and Case, *Mol. Model. Nucleic Acids* **682**, 379 (1998).

DNA bending in nature



Nucleosome (1AOI)
25°/bp



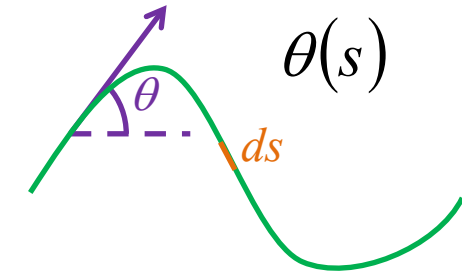
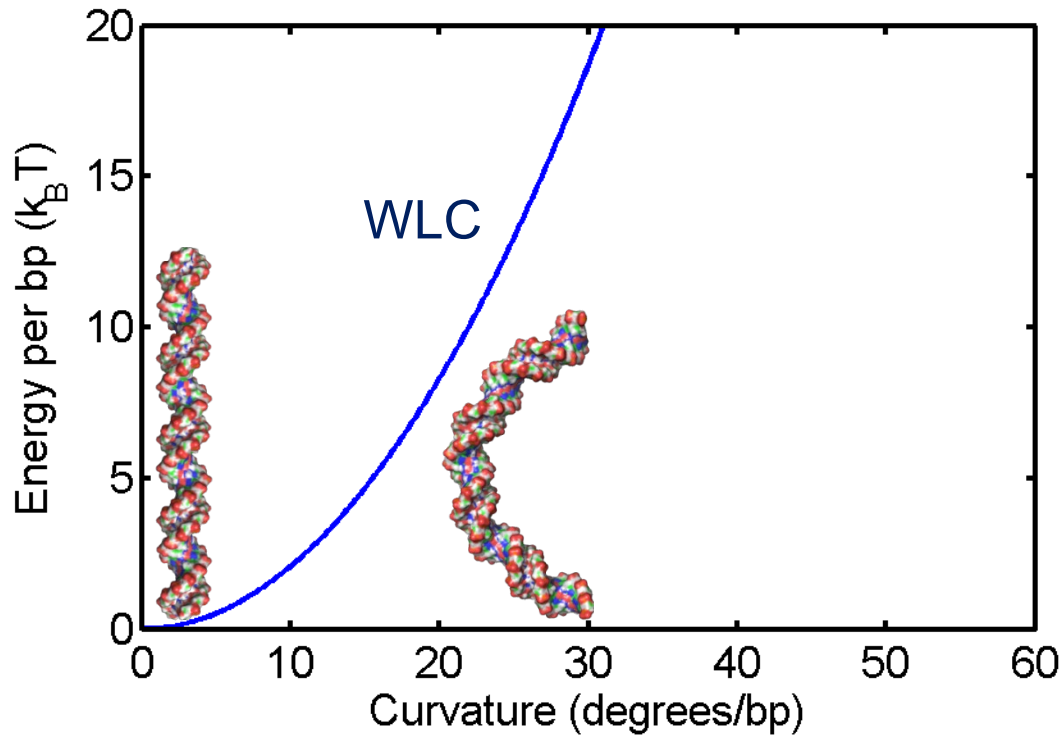
MutS + G-T mismatch (1E3M)
60°/bp at mismatch



Catabolite activator protein (1CGP)
40°/bp

Redundancy of codons allows for an underlying mechanical code

The wormlike chain model of DNA bending



Wormlike chain (WLC):

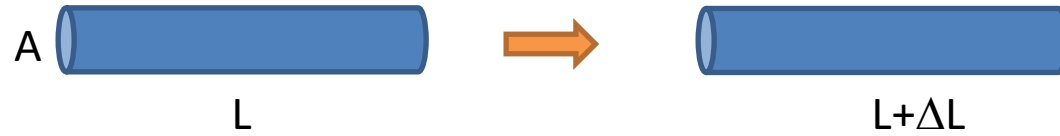
$$H = \int_0^L \frac{1}{2} \alpha \left(\frac{\partial \theta}{\partial s} \right)^2 ds$$

↓
bending modulus

- Problems with using the WLC to describe DNA bending in the context of protein binding
 - WLC treats DNA as mechanically homogeneous, but interactions reflect local variation (sequence, damage, modifications)
 - Proteins induce higher curvature than is experimentally accessible
- Idea: induce curvature in short pieces of DNA

Bending of a rod

Appetizer: **stretching**



Spring constant $k = Y \frac{A}{L}$ ↗ Young's modulus

$$\text{Elastic energy } U = \frac{1}{2} Y \frac{A}{L} \Delta L^2 = \frac{1}{2} Y A \int_0^L \left(\frac{\Delta L}{L} \right)^2 dL$$

Main course: **bending**

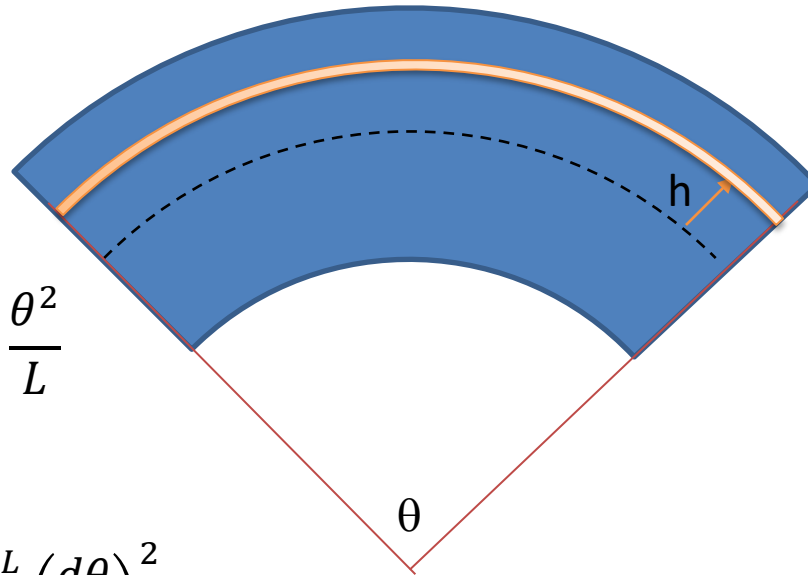
$$\Delta L = h\theta$$

$$dU = \frac{1}{2} Y \frac{dA}{L} h^2 \theta^2$$

$$U = \frac{1}{2} Y \left(\int_{surf} h^2 dA \right) \frac{\theta^2}{L}$$

⏟
 I

$$U = \frac{1}{2} Y I \frac{\theta^2}{L} = \frac{1}{2} Y I \int_0^L \left(\frac{d\theta}{dL} \right)^2 ds$$



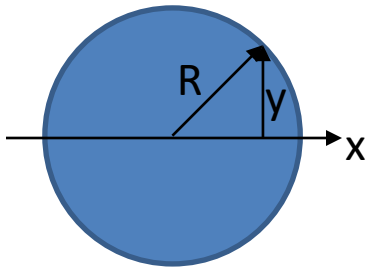
Relation of bending modulus to material properties

$$\alpha = Y I$$

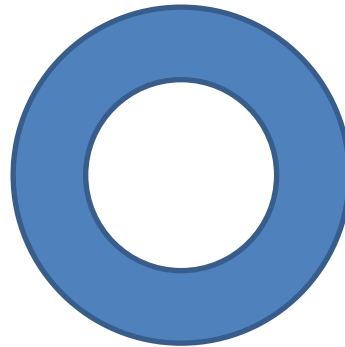
Area moment of inertia (m^4)

Young's Modulus (N/m^2)

$$I = \int_A y^2 dA$$



$$I_x = \frac{\pi R^4}{4}$$



$$I = \frac{\pi(R_2^4 - R_1^4)}{4}$$

(easiest to see by calculating I_z and then applying perpendicular axis theorem)

Why are microtubules hollow?

Coherent bending ($I \sim R^4$) vs. incoherent bending ($I \sim R^2$)

Mechanics of rods

$$H = \int_0^L \frac{1}{2} \alpha \left(\frac{\partial \theta}{\partial s} \right)^2 ds$$

Loop energy: $\frac{\partial \theta}{\partial s} = \frac{1}{R}$

$$U_{loop} = 2\pi R \frac{1}{2} \alpha \left(\frac{1}{R} \right)^2$$
$$= \frac{\pi \alpha}{R}$$

Consider $U_{loop} = \frac{1}{2} k_B T$

$$\rightarrow R = 2\pi \frac{\alpha}{k_B T}$$

Persistence length $l_p = \frac{\alpha}{k_B T}$

e.g. for dsDNA, $l_p \sim 53$ nm

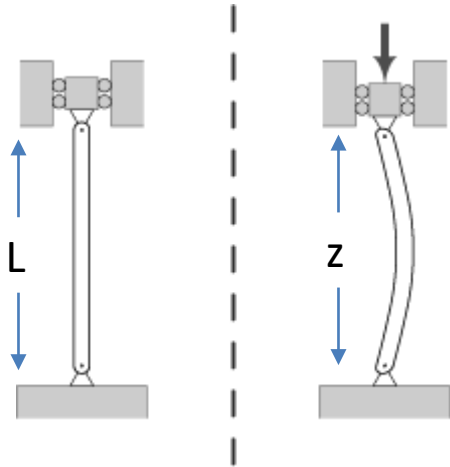
Autocorrelation of unit tangent $\langle \mathbf{t}(s_a) \cdot \mathbf{t}(s_b) \rangle = e^{-|s_a - s_b|/l_p}$

Example: DNA bending around a histone

Histone: $R \sim 4$ nm, 2 revolutions

$U_{bend} \sim 70 k_B T$

Rods with forces: Euler buckling



$$U_{tot} = \frac{1}{2} \alpha \int_0^L \left(\frac{d\theta}{ds} \right)^2 ds - F(L - z)$$

Guess: $\theta(s) = A \cos \frac{\pi s}{L}$

$$\left(\frac{d\theta}{ds} \right)^2 = \left(\frac{A\pi}{L} \sin \frac{\pi s}{L} \right)^2$$

$$U_{bend} = \frac{1}{2} \alpha \left(A \frac{\pi}{L} \right)^2 \frac{L}{2}$$

$$= \frac{\alpha A^2 \pi^2}{4L}$$

$$z = \int_0^L \cos \theta ds$$

$$= \int_0^L \cos \left(A \cos \frac{\pi s}{L} \right) ds$$

$$\sim \int_0^L \left[1 - \frac{1}{2} \left(A \cos \frac{\pi s}{L} \right)^2 \right] ds$$

$$= L - \frac{A^2 L}{4}$$

$$U_{mech} = -\frac{FA^2 L}{4}$$

$$U_{tot} = A^2 \left(\frac{\alpha \pi^2}{4L} - \frac{FL}{4} \right)$$

$$F_{buckle} \geq \frac{\pi^2 \alpha}{L^2} \quad L_{buckle} \geq \sqrt{\frac{\pi^2 \alpha}{F}}$$

The elasticae

For a rod under compression $U(s) = \frac{\alpha}{2} \left(\frac{\partial \theta}{\partial s} \right)^2 + F \cos \theta$

Lagrangian of a physical pendulum

$$\begin{aligned} \mathcal{L} &= KE - PE \\ &= \frac{1}{2} mL^2 \left(\frac{d\theta}{dt} \right)^2 + mgL \cos \theta \end{aligned}$$

Lagrangian is minimized via
Calculus of Variations

$$\frac{d}{dt} \left(\frac{\partial \mathcal{L}}{\partial \dot{q}} \right) = \frac{\partial \mathcal{L}}{\partial q}$$

→ Newton's eqns of motion

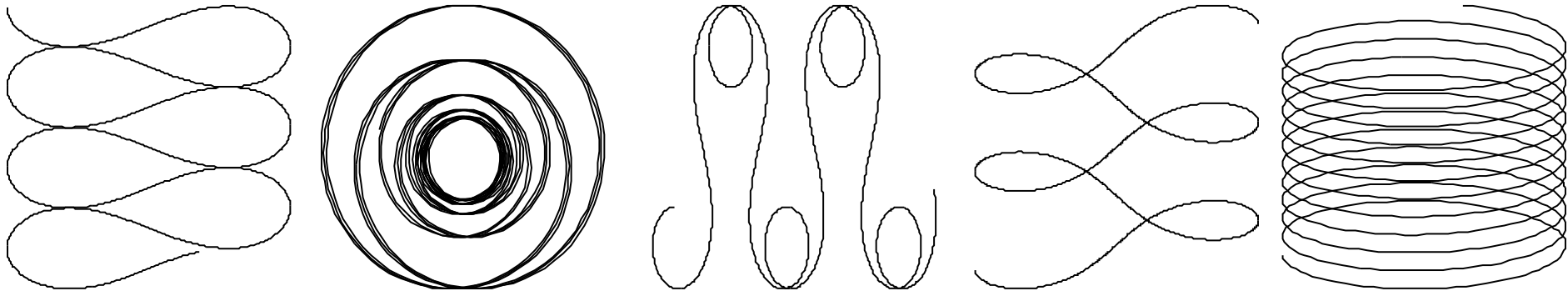
For a rod:

$$\frac{d}{ds} \alpha \left(\frac{\partial \theta}{\partial s} \right) = -F \sin \theta$$

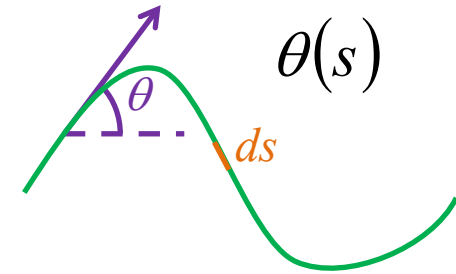
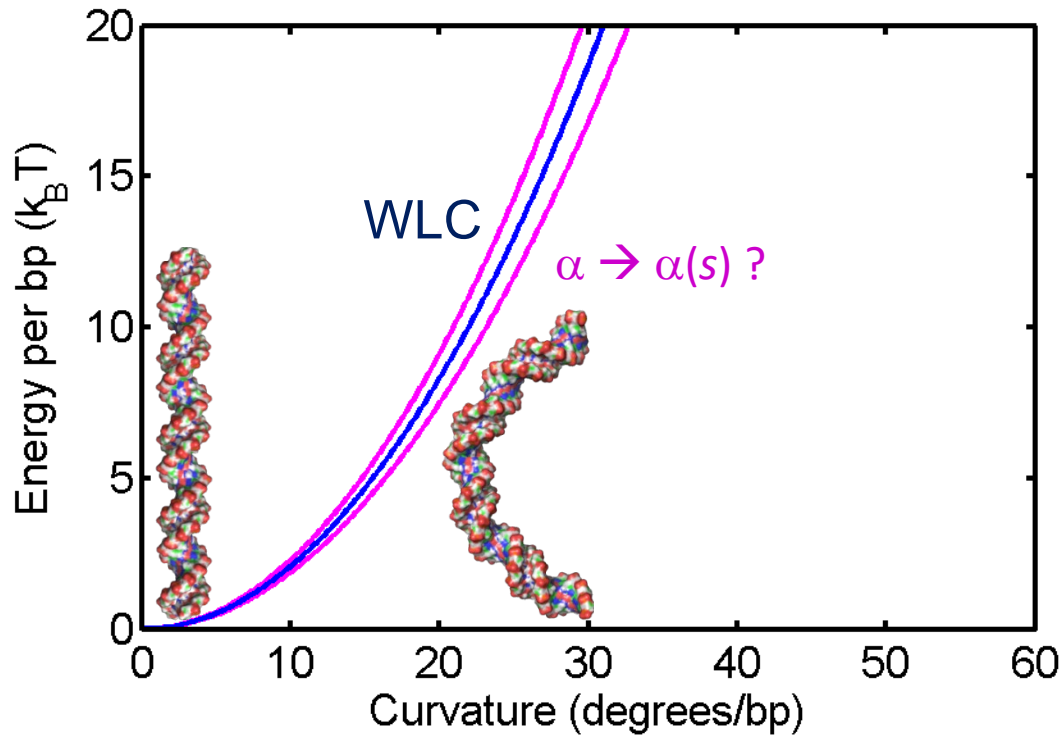
Small-angle limit:

$$\alpha \frac{d^2 \theta}{ds^2} = -F \theta$$

$$\theta(s) = A \sin \sqrt{\frac{F}{\alpha}} s + B \cos \sqrt{\frac{F}{\alpha}} s$$



The wormlike chain model of DNA bending



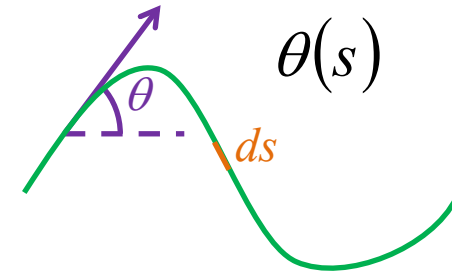
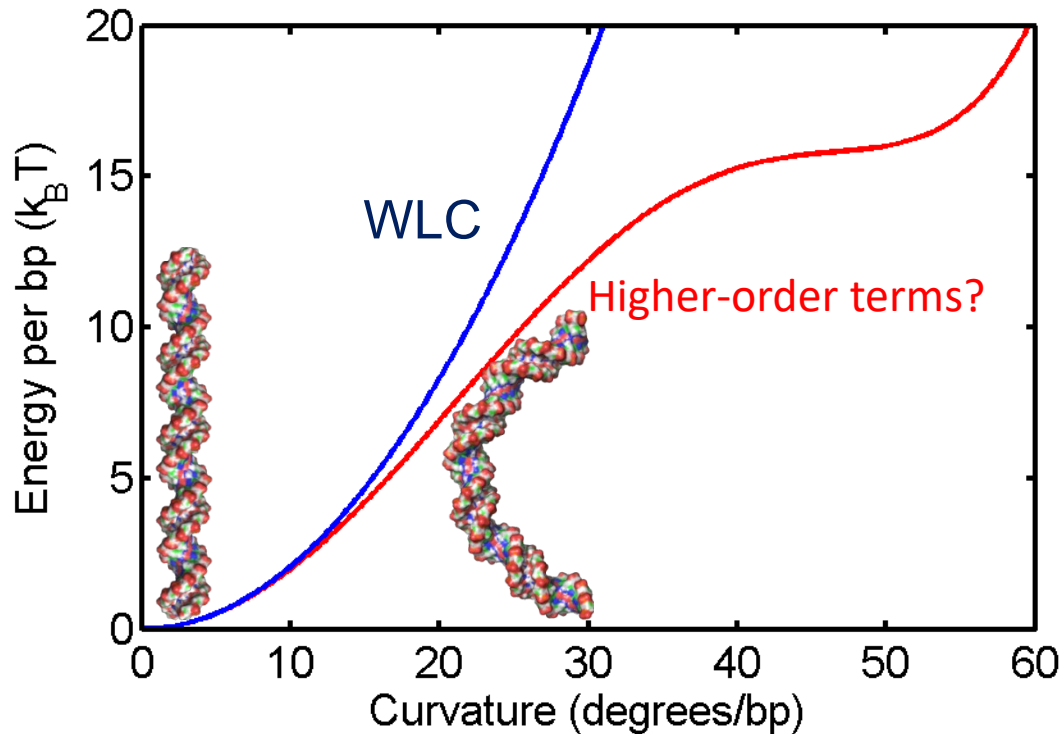
Wormlike chain (WLC):

$$H = \int_0^L \frac{1}{2} \alpha \left(\frac{\partial \theta}{\partial s} \right)^2 ds$$

↓
bending modulus

- Problems with using the WLC to describe DNA bending in the context of protein binding
 - WLC treats DNA as mechanically homogeneous, but interactions reflect local variation (sequence, damage, modifications)
 - Proteins induce higher curvature than is experimentally accessible
- Idea: induce curvature in short pieces of DNA

The wormlike chain model of DNA bending



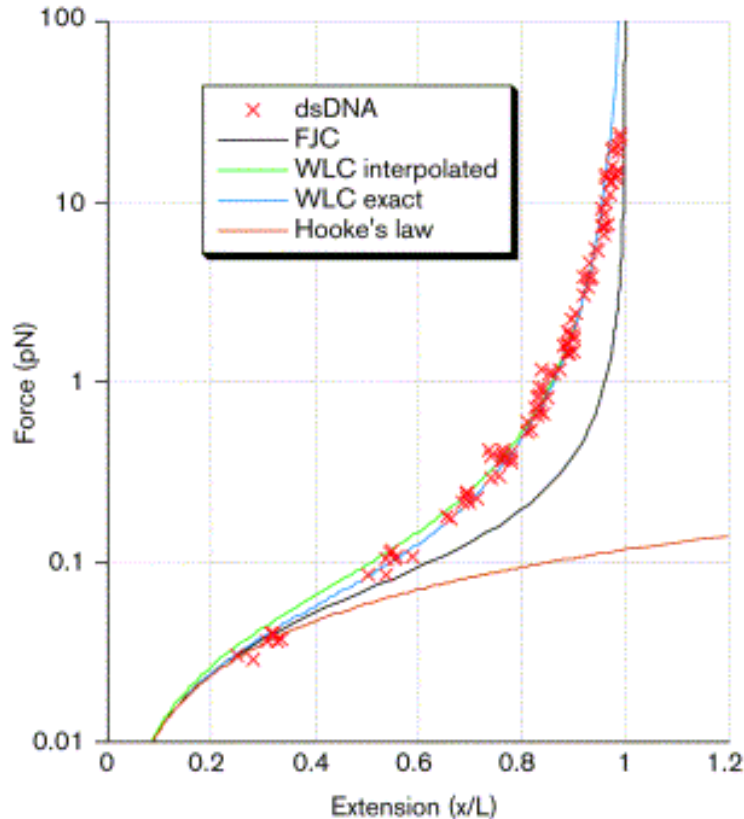
Wormlike chain (WLC):

$$H = \int_0^L \frac{1}{2} \alpha \left(\frac{\partial \theta}{\partial s} \right)^2 ds$$

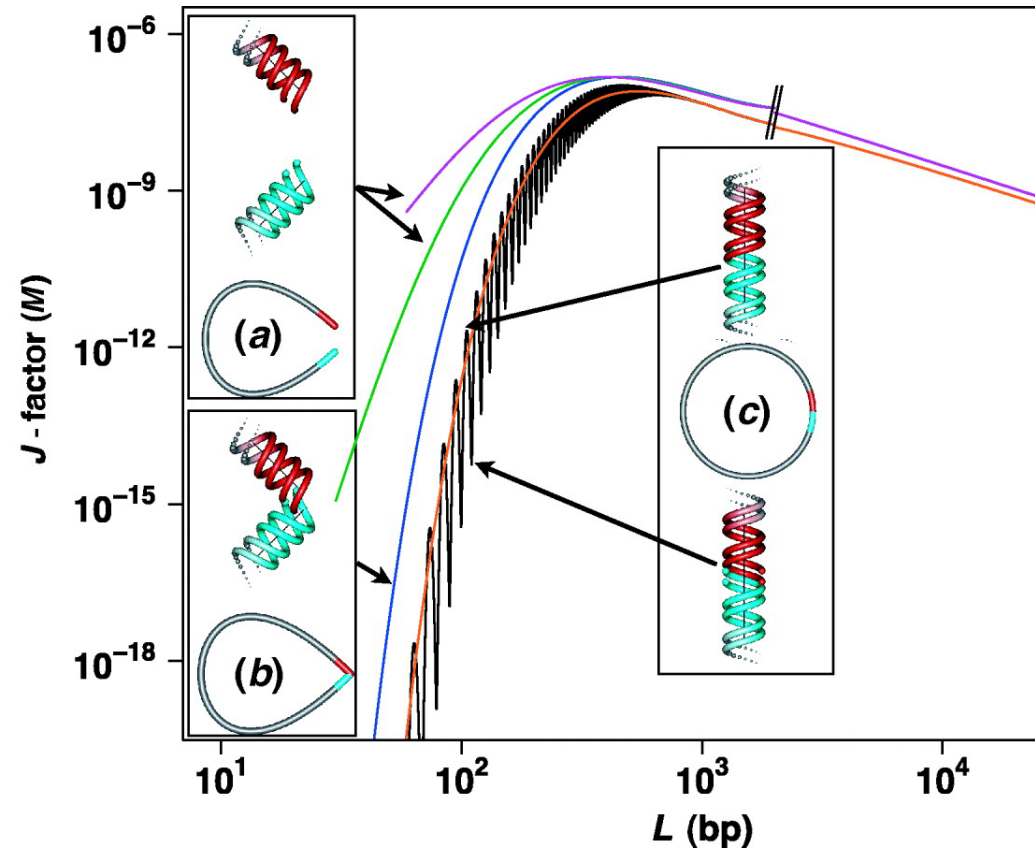
↓
bending modulus

- Problems with using the WLC to describe DNA bending in the context of protein binding
 - WLC treats DNA as mechanically homogeneous, but interactions reflect local variation (sequence, damage, modifications)
 - Proteins induce higher curvature than is experimentally accessible
- Idea: induce curvature in short pieces of DNA

Experiments consistent with elastic rod model



Bustamante *et al.* (2000), *Curr. Opin. Struct. Biol.*

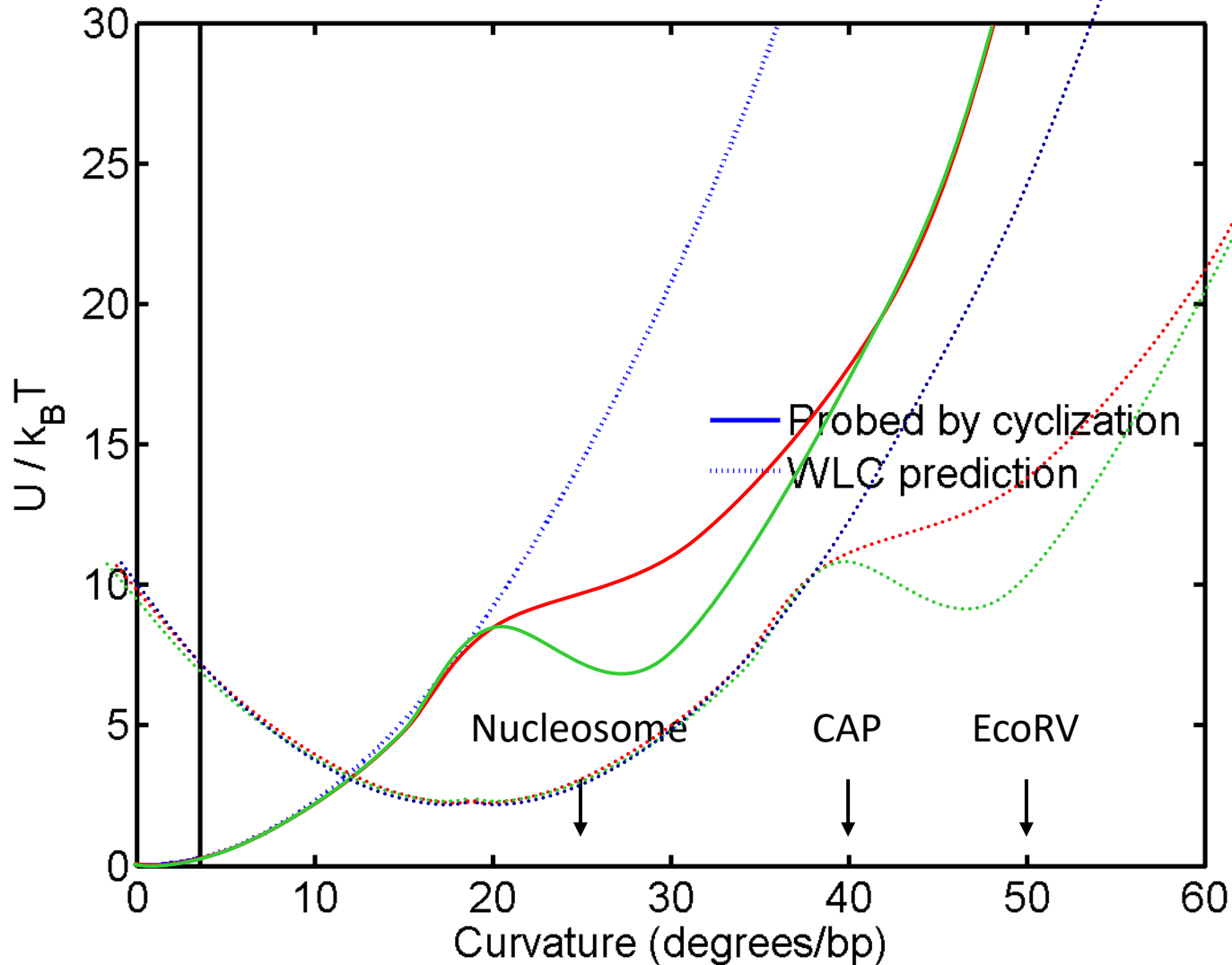


Peters and Maher (2010), *Quarterly Rev. of Biophys.*

WLC model is powerful because it is insensitive to molecular details

but sometimes details matter...

Current experiments are limited to thermally accessible bending energy



Where might the homogeneous elastic rod model fail?

- **Particular sequences**

Nucleosome positioning sequences; protein binding sites

- **Regions of high imposed curvature**

Under bound proteins; in a virus capsid; regulatory loops

- **Damage sites**

Thymine dimers, oxidation, alkylation, hydrolysis, deamination

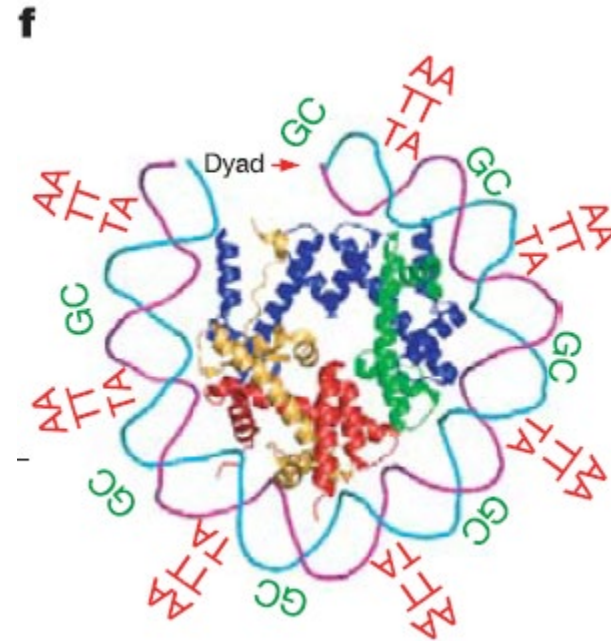
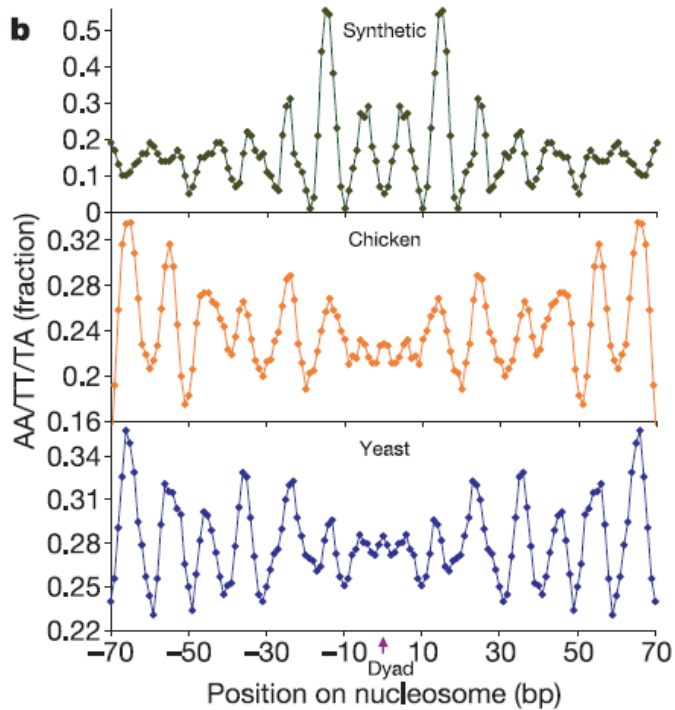
- **Base pair mismatches**

- **Nicks**

- **Gaps**

- **Epigenetic modifications**

Nucleosome positioning sequences

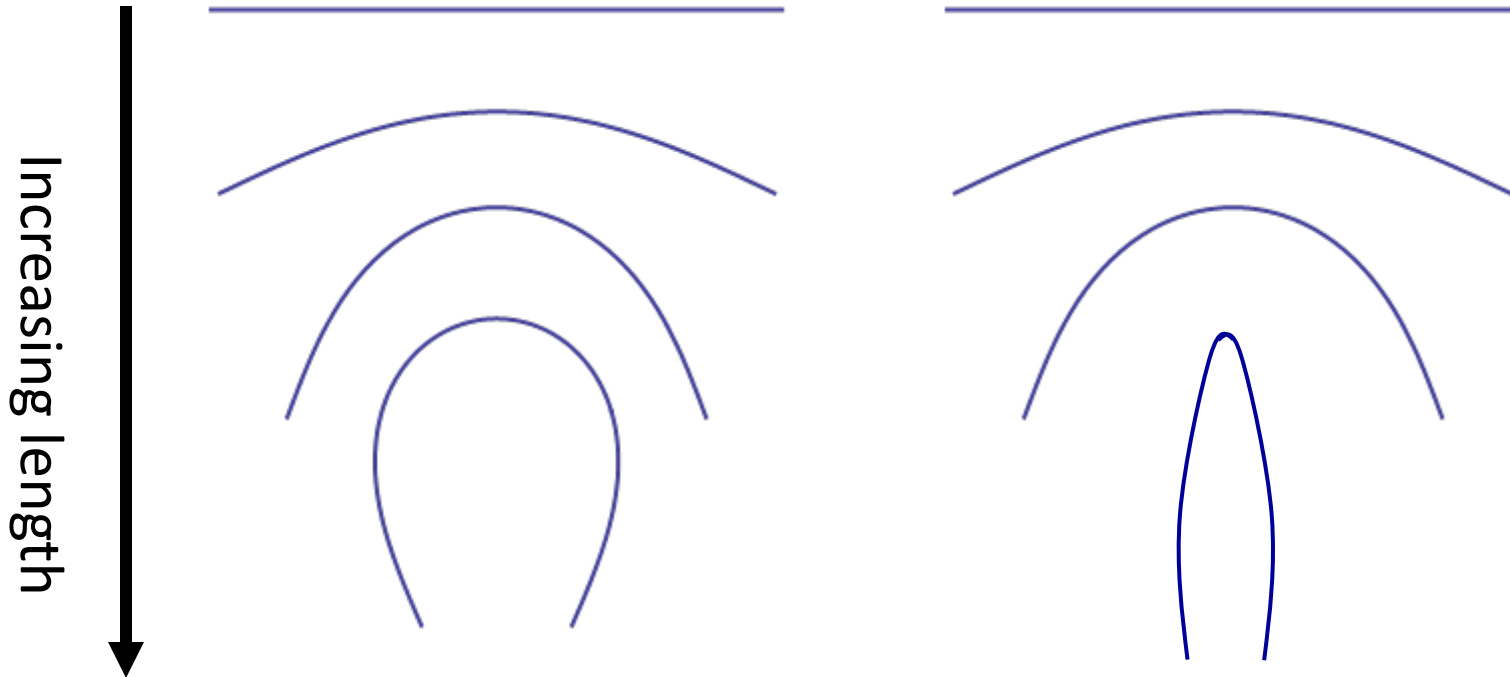


Segal, Eran, et al. "A genomic code for nucleosome positioning." *Nature* 442.7104 (2006): 772-778.

Behavior beyond buckling

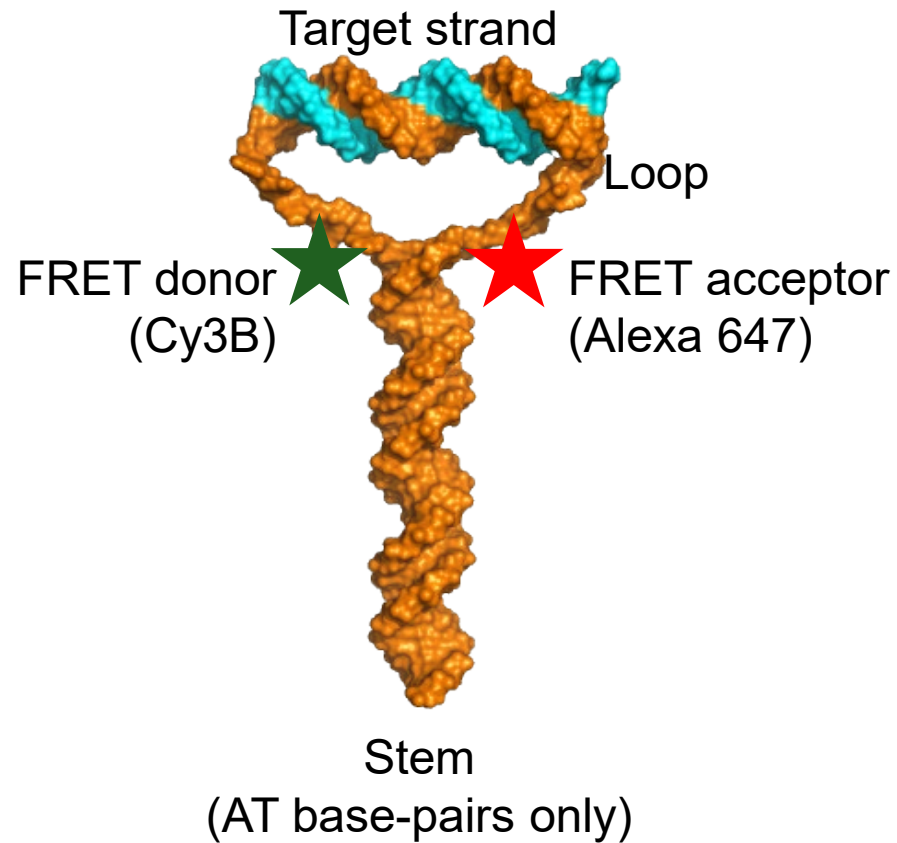
Buckling
(linear elasticity)

Kinking
(nonlinear elasticity)

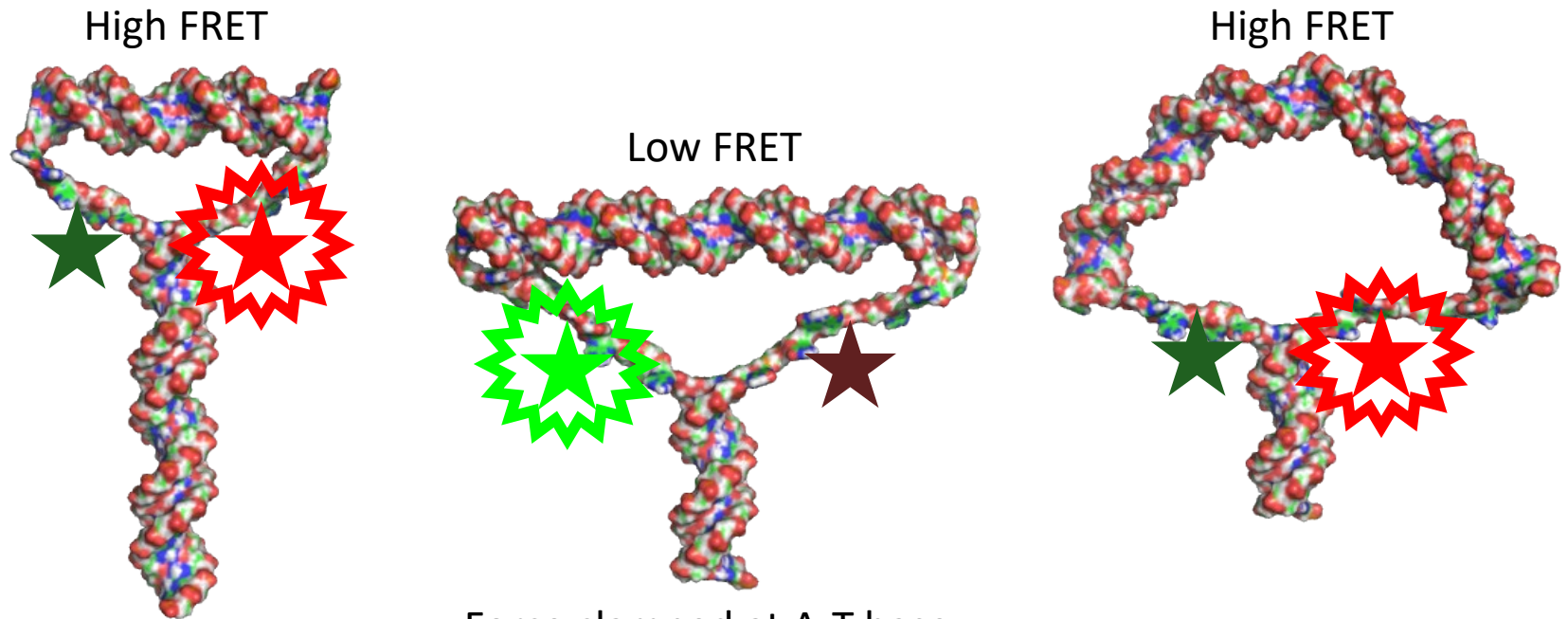


Constant compressive force

Molecular vise



Increasing complement length \longrightarrow



Force clamped at A-T base-pairing force (9 pN [1])

$$F_{AT} \approx 9 \text{ pN [1]}$$

$$\text{persistence length} \equiv \frac{\alpha}{k_B T} \approx 44 - 55 \text{ nm [2]}$$

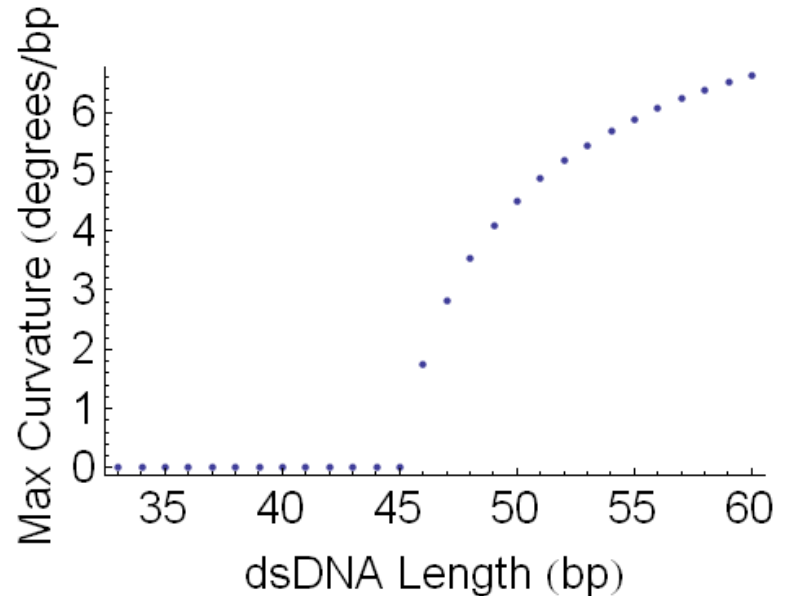
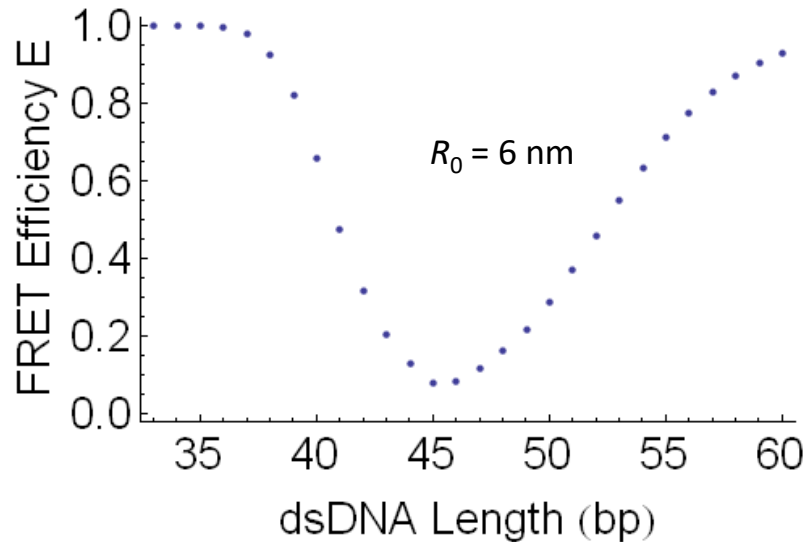
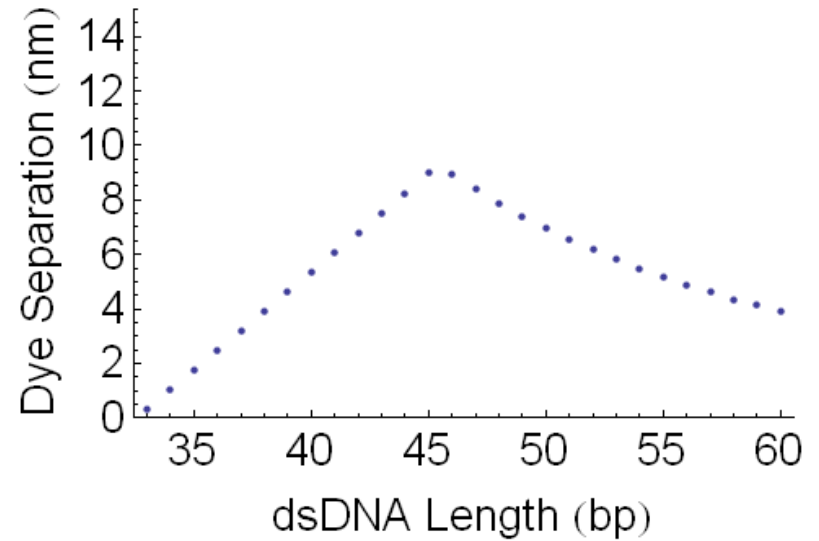
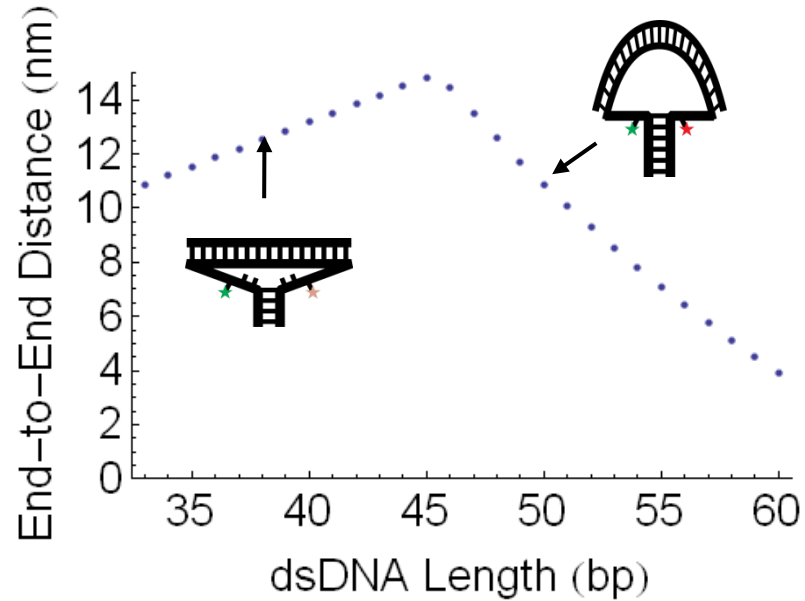
$$L_{buckle} = \sqrt{\frac{\pi^2 \alpha}{F}} \approx 14.1 - 15.7 \text{ nm} = 41 - 46 \text{ bp}$$

Stem composition	Buckling length	
A-T (9 pN)	45 bp	15 nm
G-C (21 pN)	29 bp	10 nm

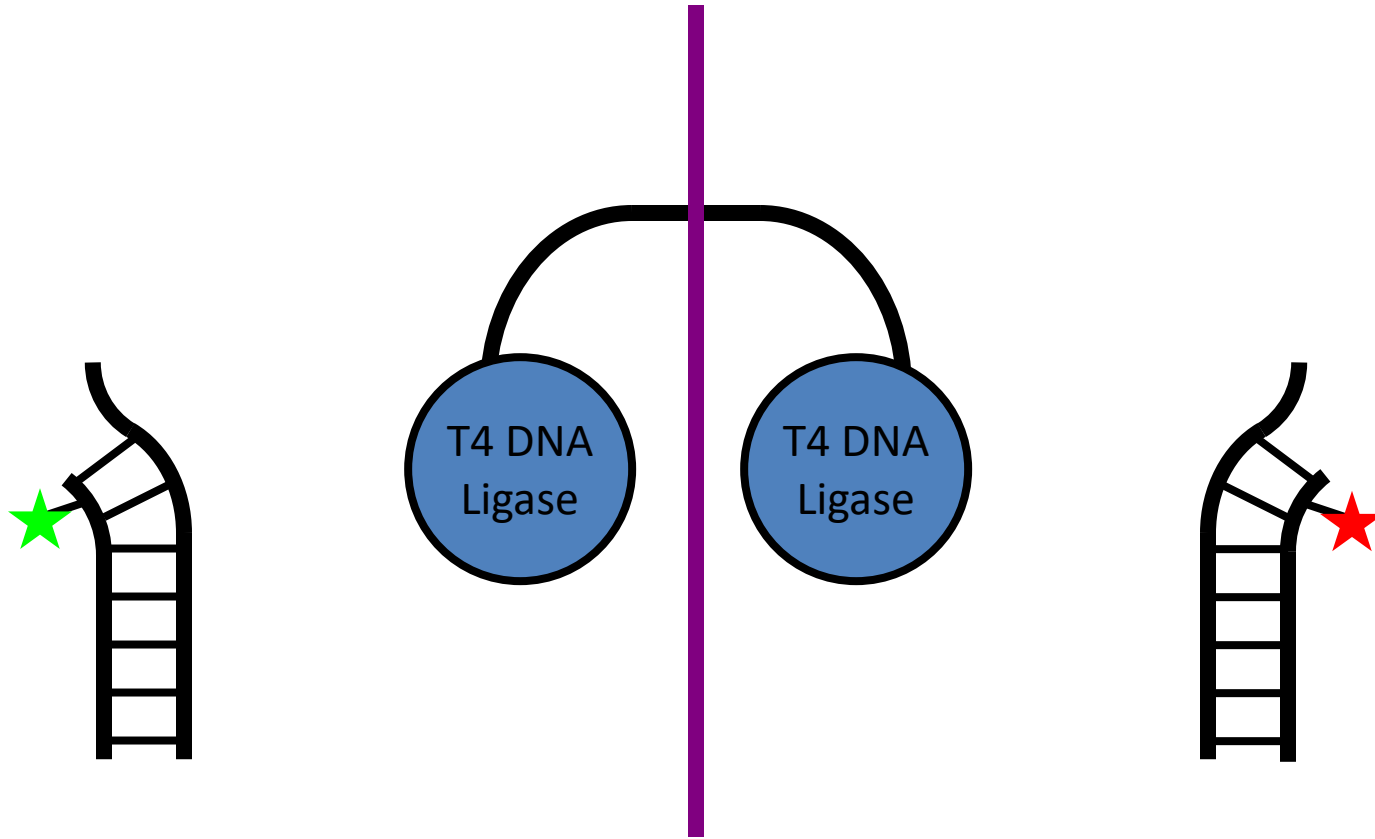
1. Woodside *et al.*, *PNAS* **103**, 6190 (2006).

2. Lu, Weers, Stellwagen, *Biopolymers* **61**, 261 (2002).

Predictions of linear elasticity



Hairpin Synthesis



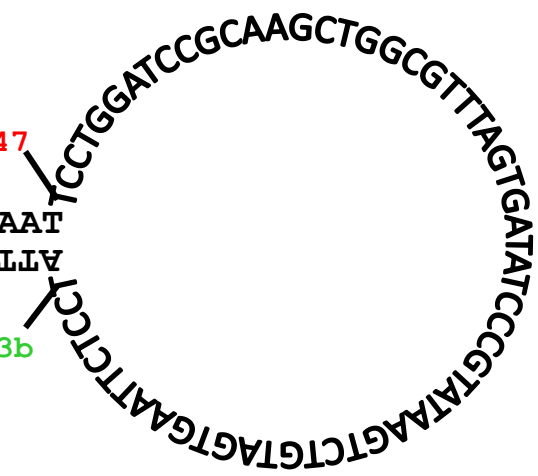
Loop: 60 nt

Short Stem: G-C 2; A-T 33

5' - GCAATTTATTAATTATATATTTTATTTAATATAAT
3' - CTTAATTTAATTTATATATTTTATTTAATATAAT

Alexa 647

Cy3b

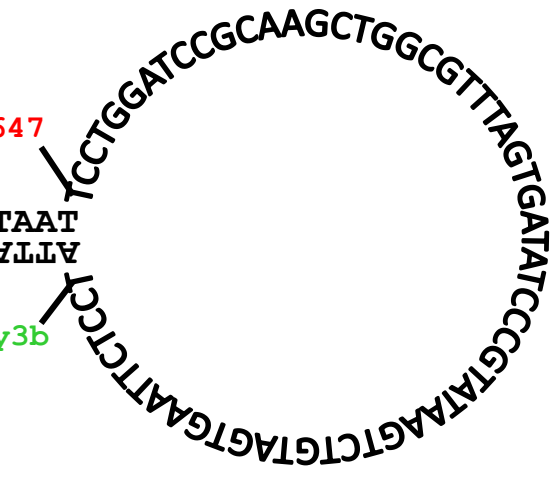


Long Stem: G-C 10; A-T 39

5' - GCCCGGCGGCTTATAAAATTTATTAATTATATATTTTATTTAATATAAT
3' - CTTAATTTAATTTATATATTTTATTTAATATAAT

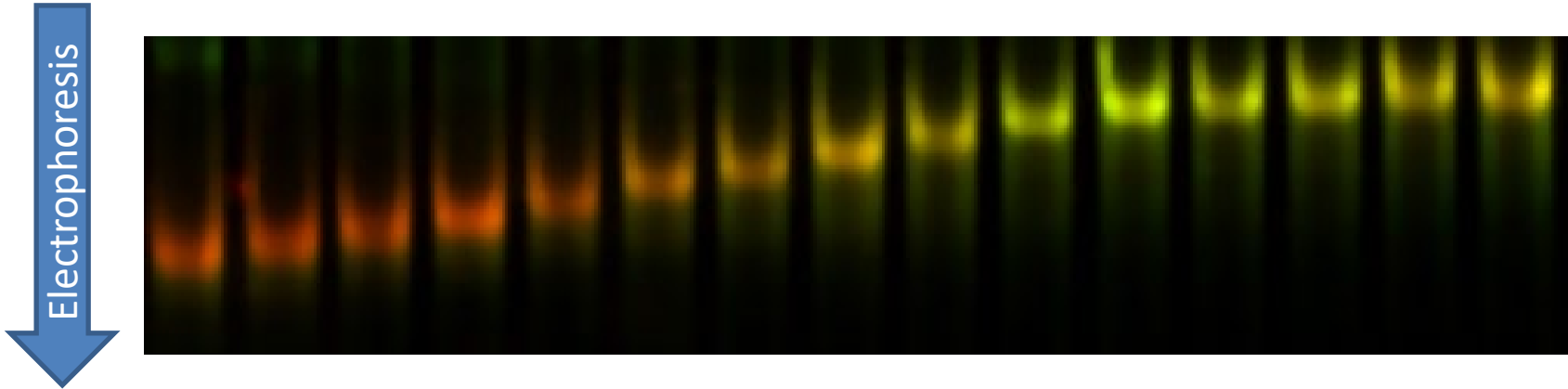
Alexa 647

Cy3b

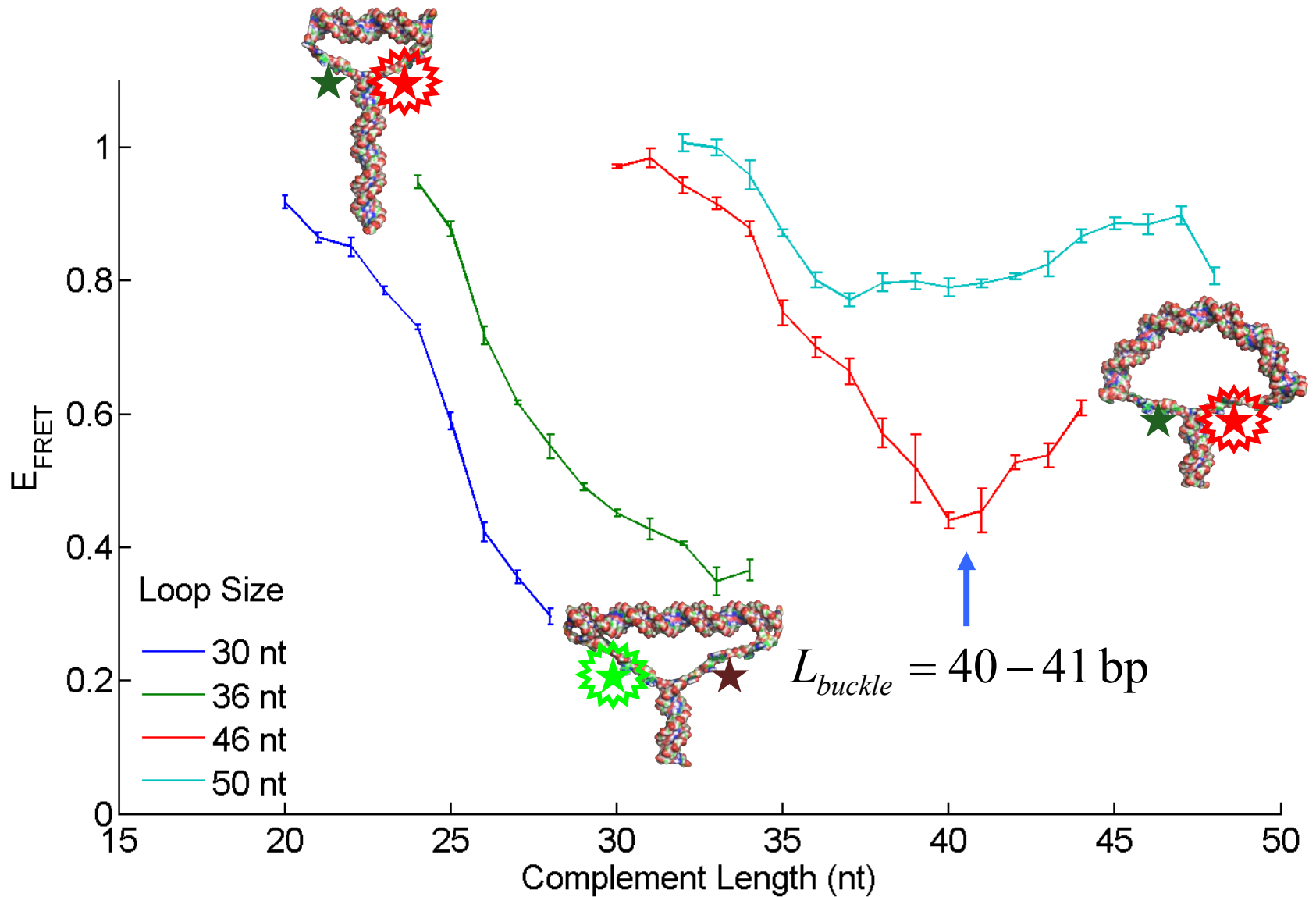


Data collection: native PAGE

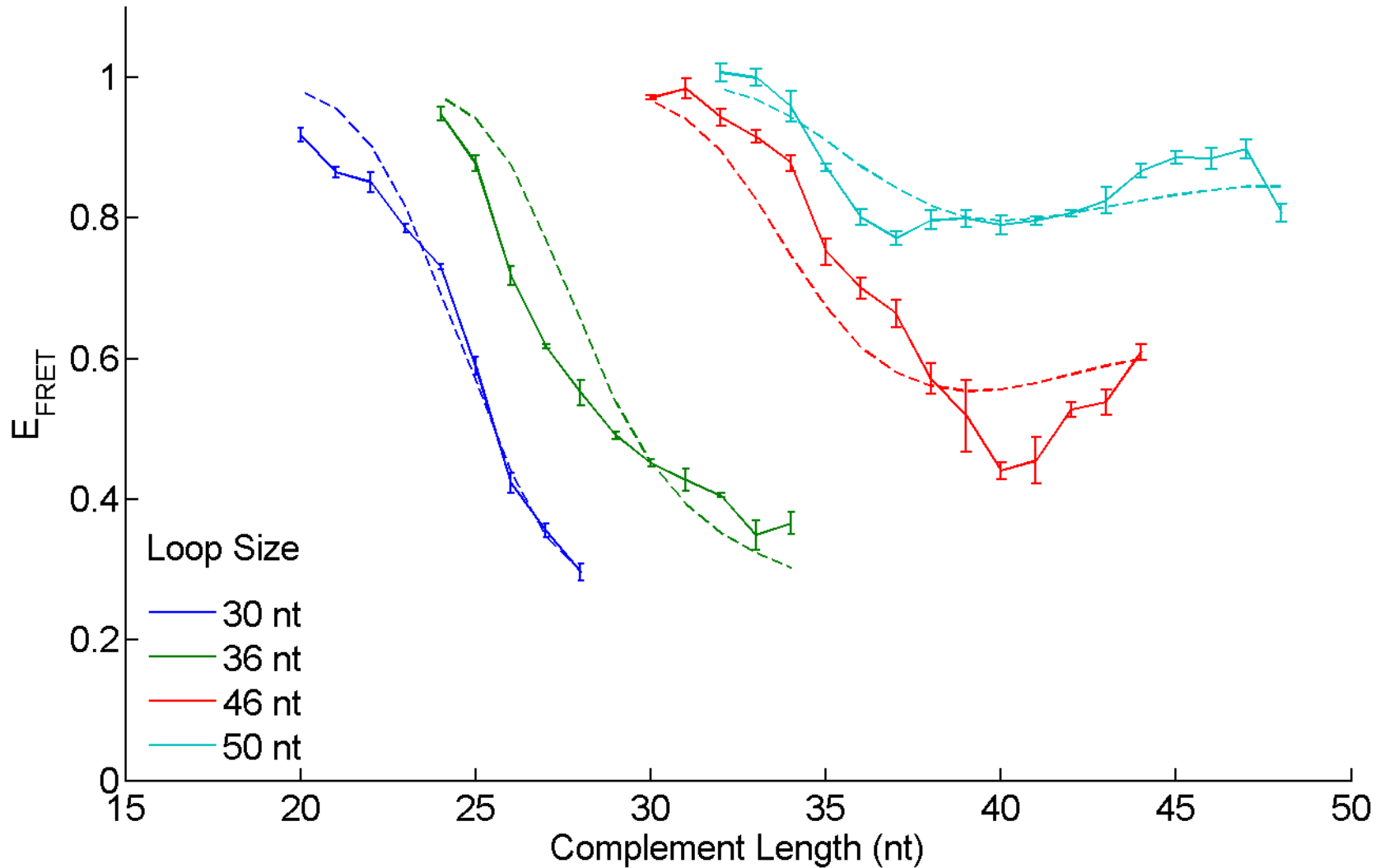
Increasing complement length →



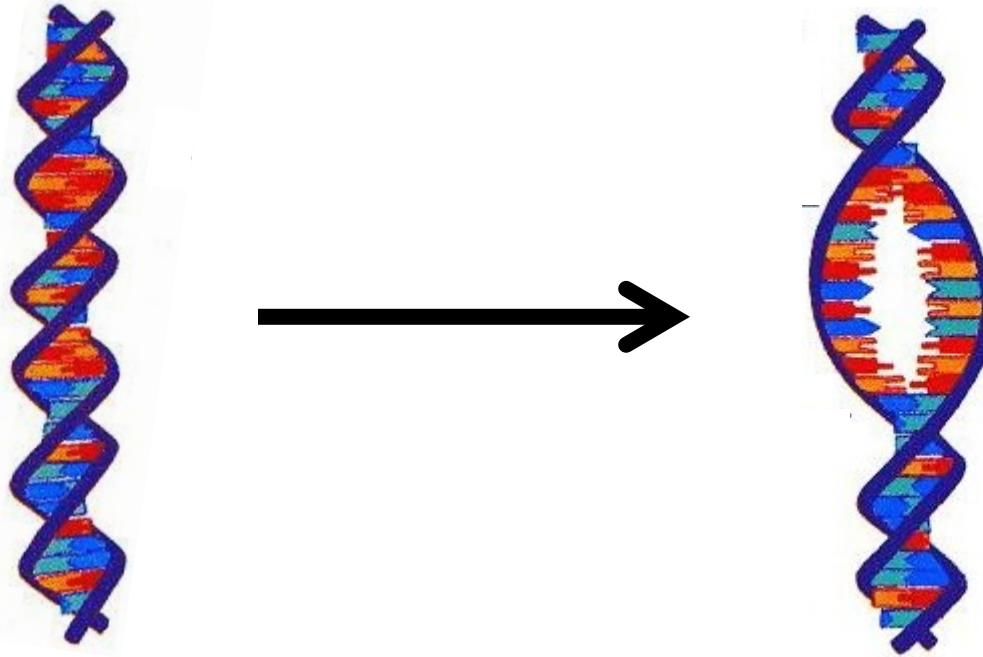
Euler buckling in dsDNA



A simple statistical mechanical model

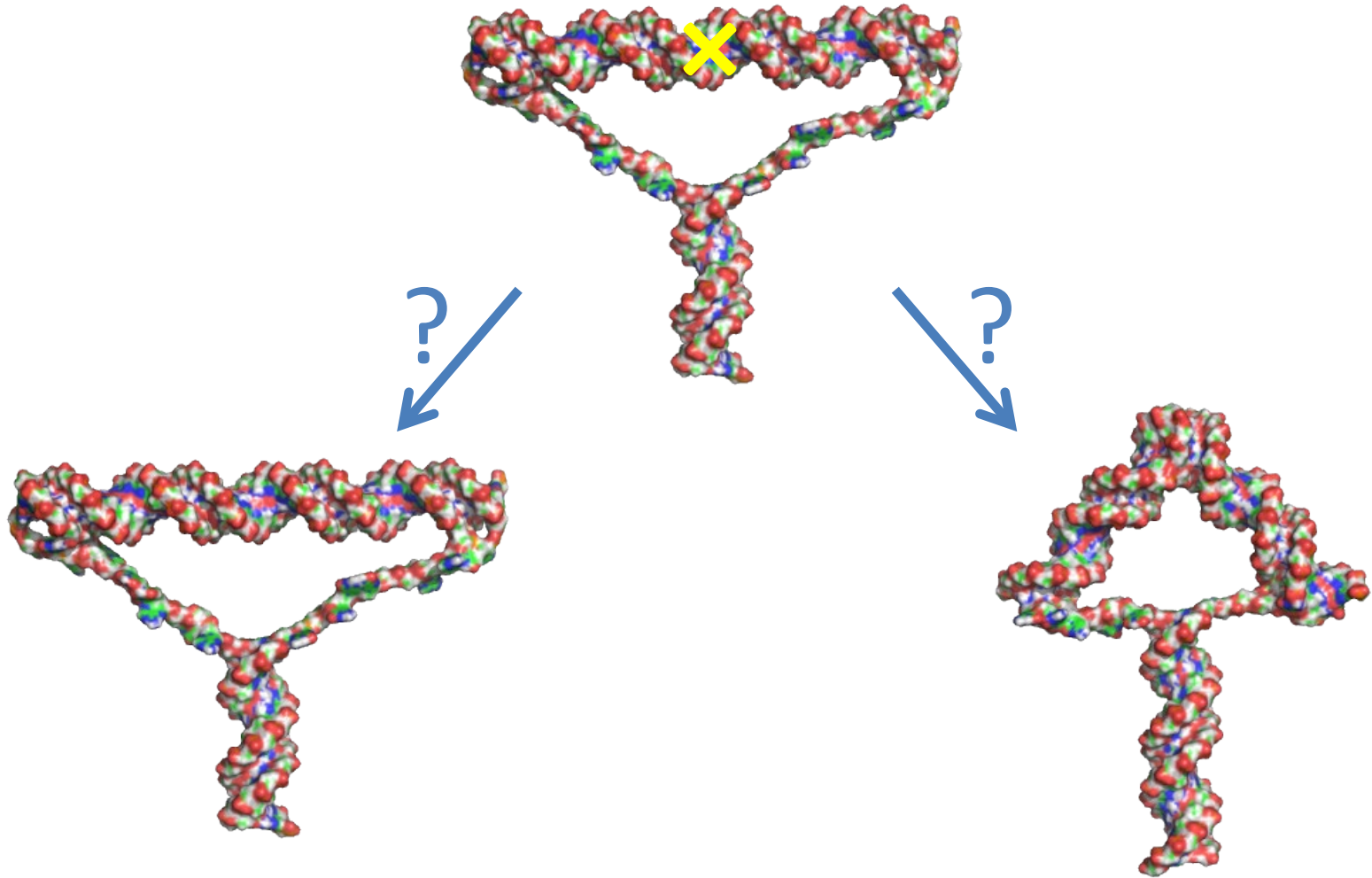


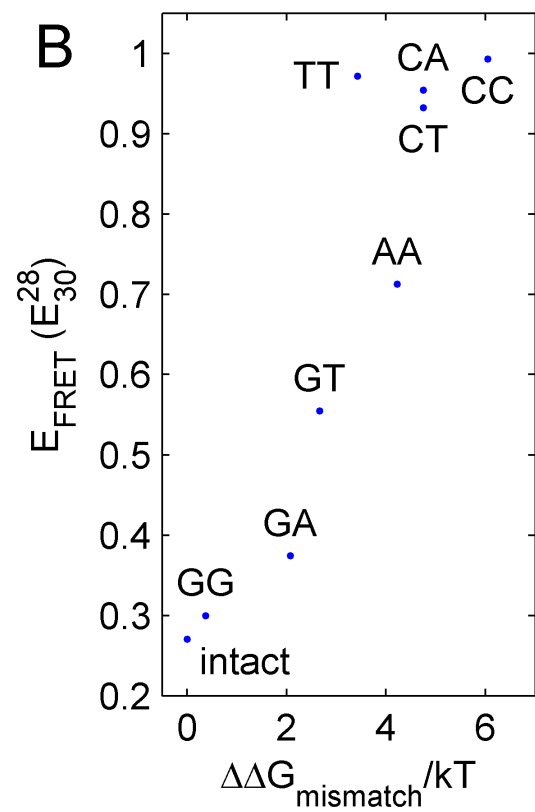
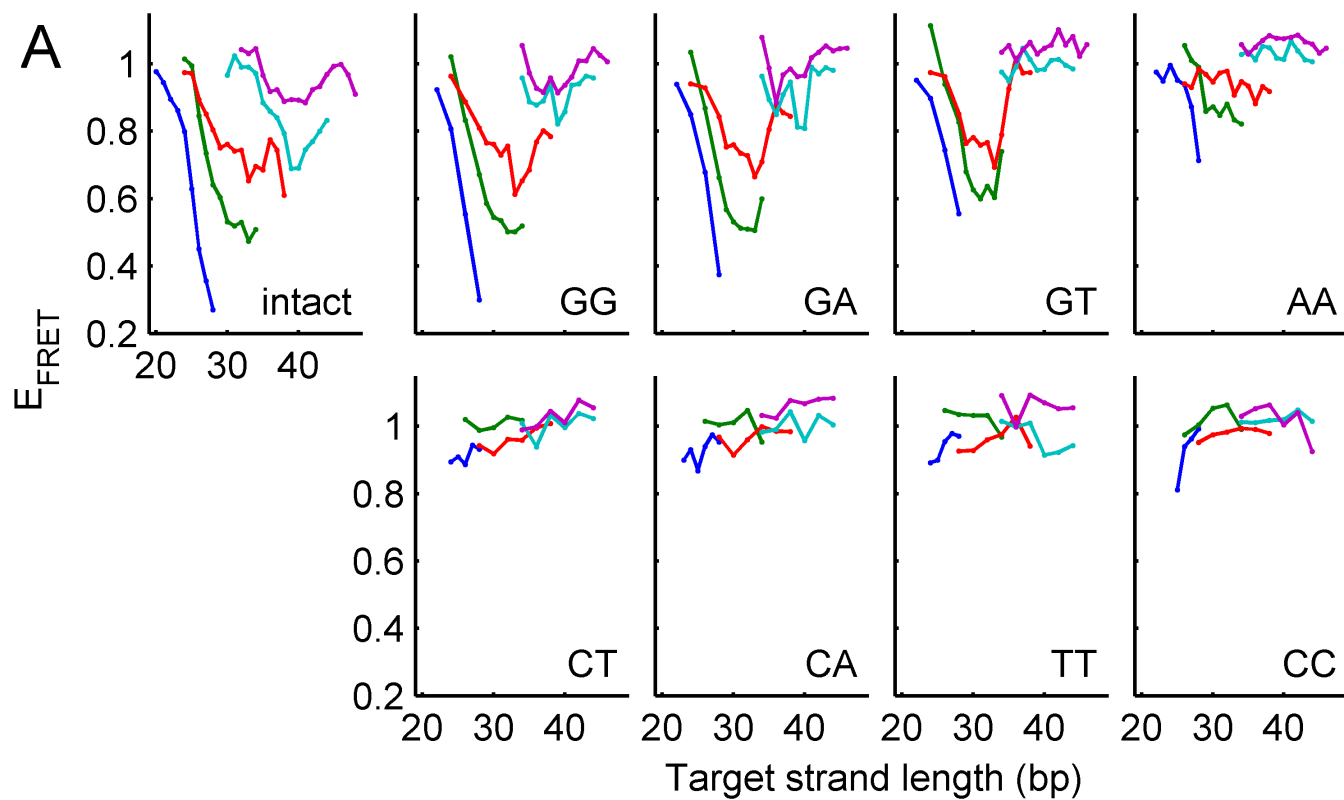
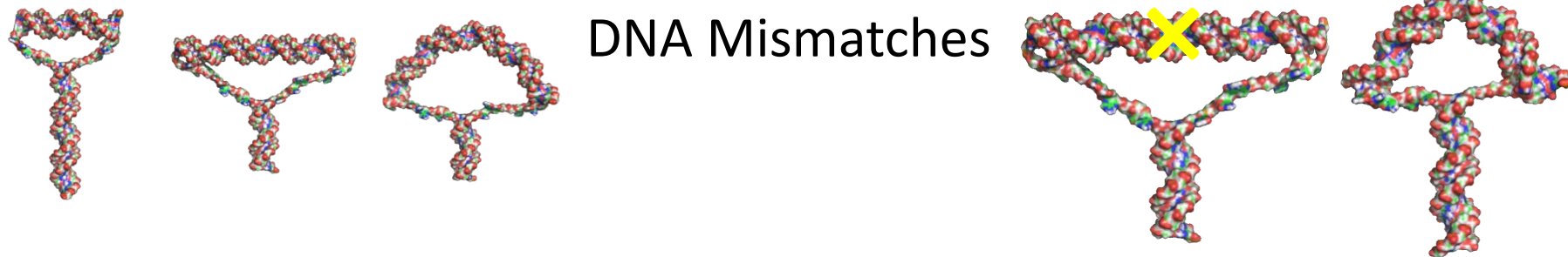
Kinking by local melting



- Hypotheses:
 - Kinking should correlate with “meltability”
 - Kinked structures should be recognizable as single-stranded DNA

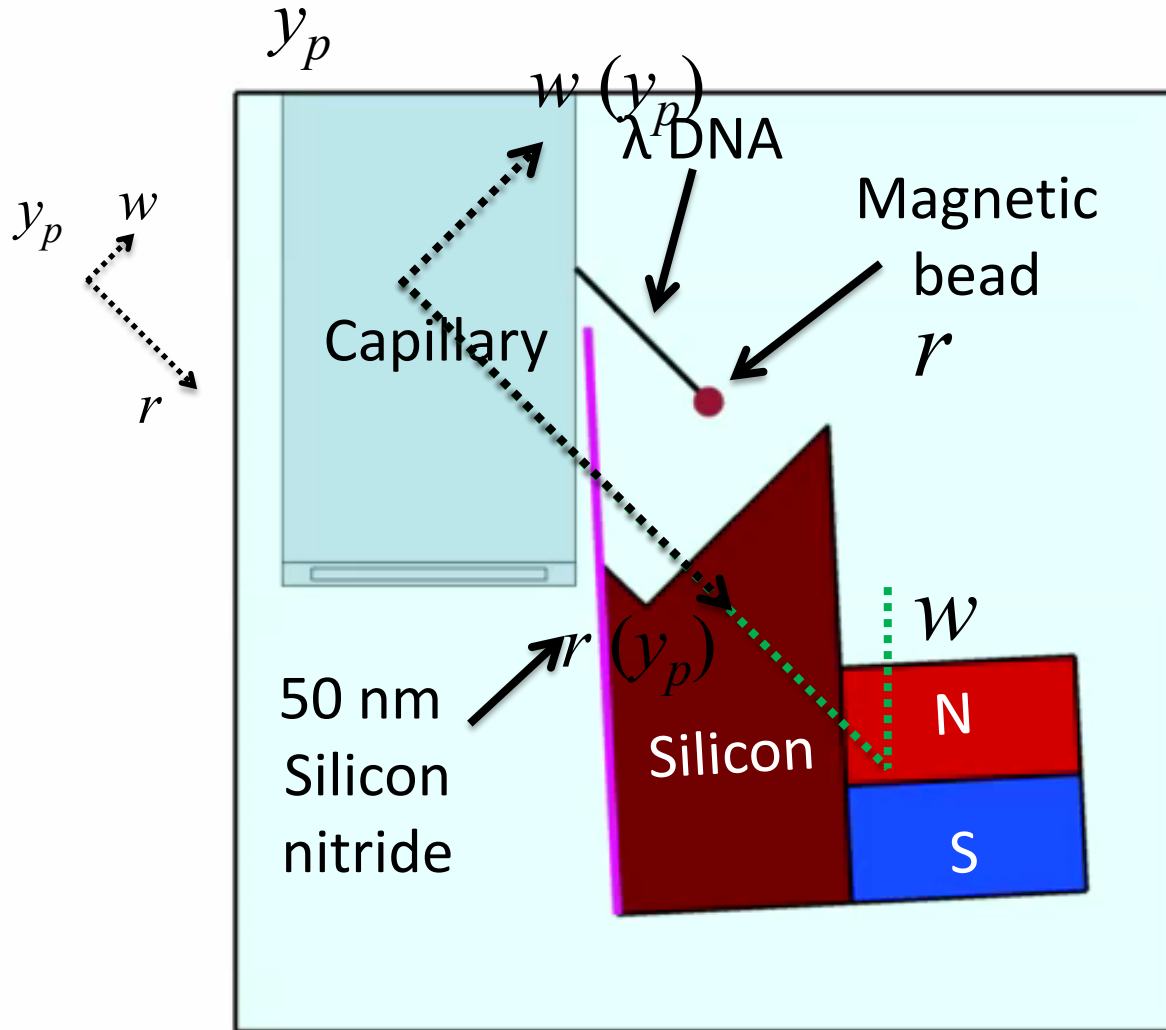
DNA Mismatches



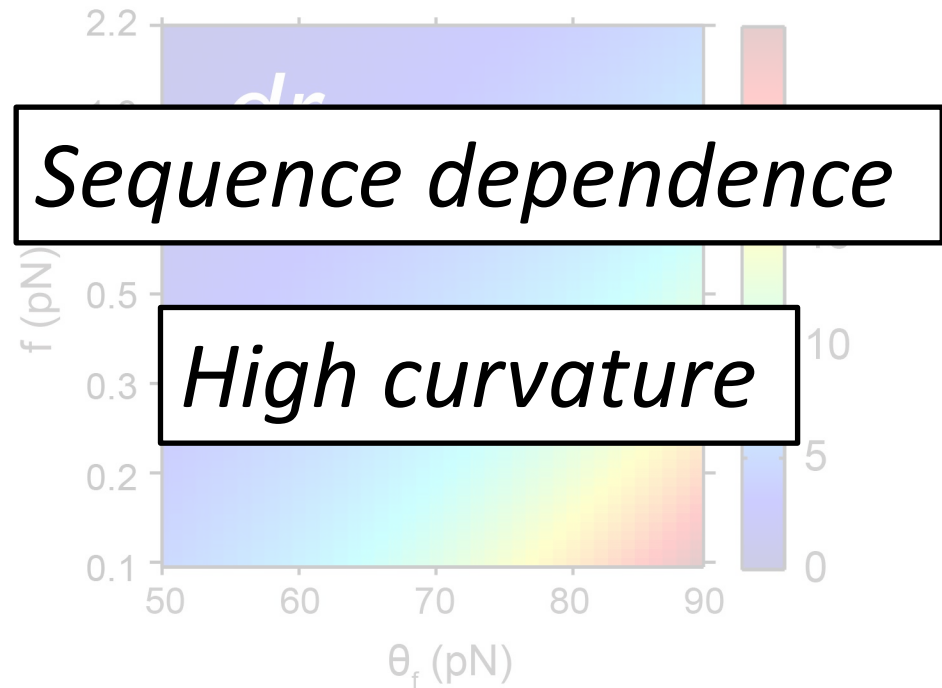
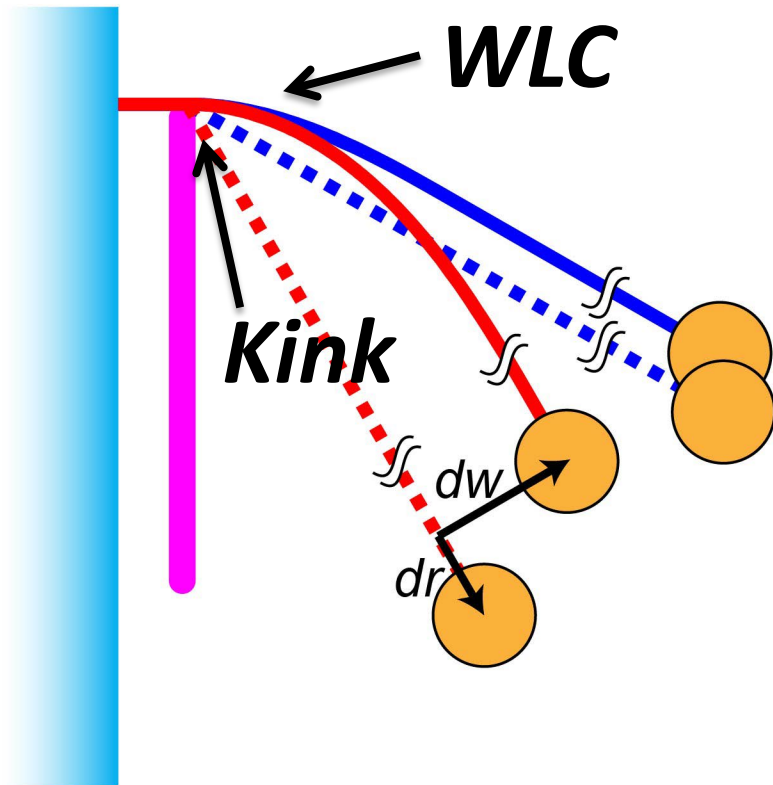


Spearman's $\rho = 0.90$
($P = 0.0022$)

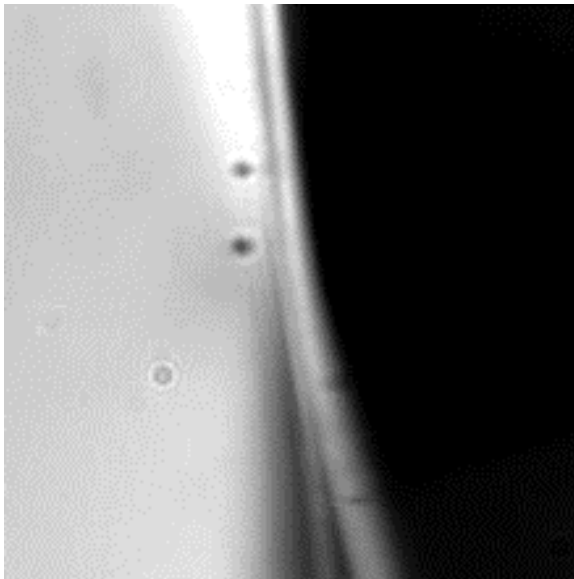
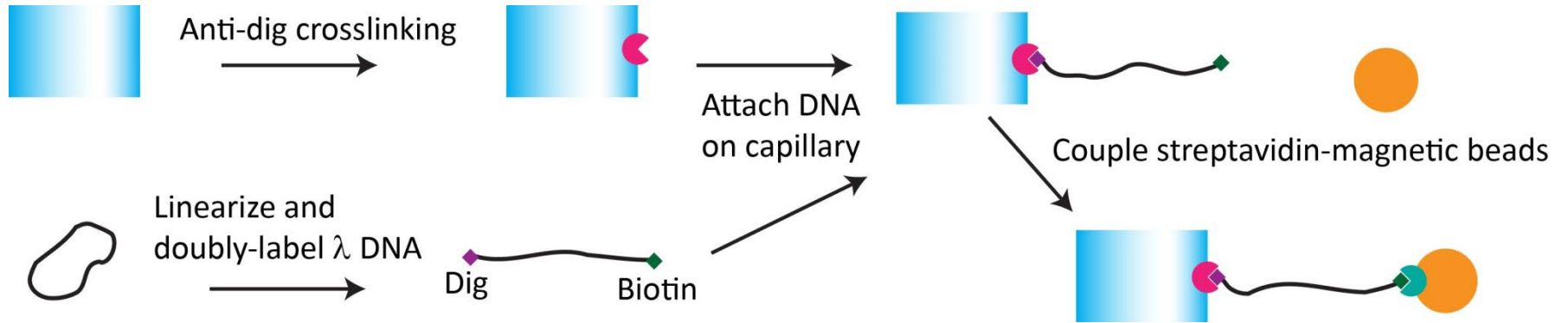
DNA Pulley



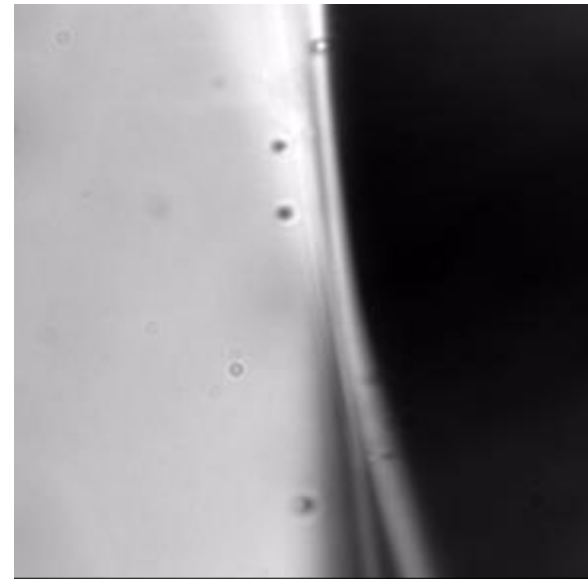
Nano-mechanics of DNA



λ DNA construct

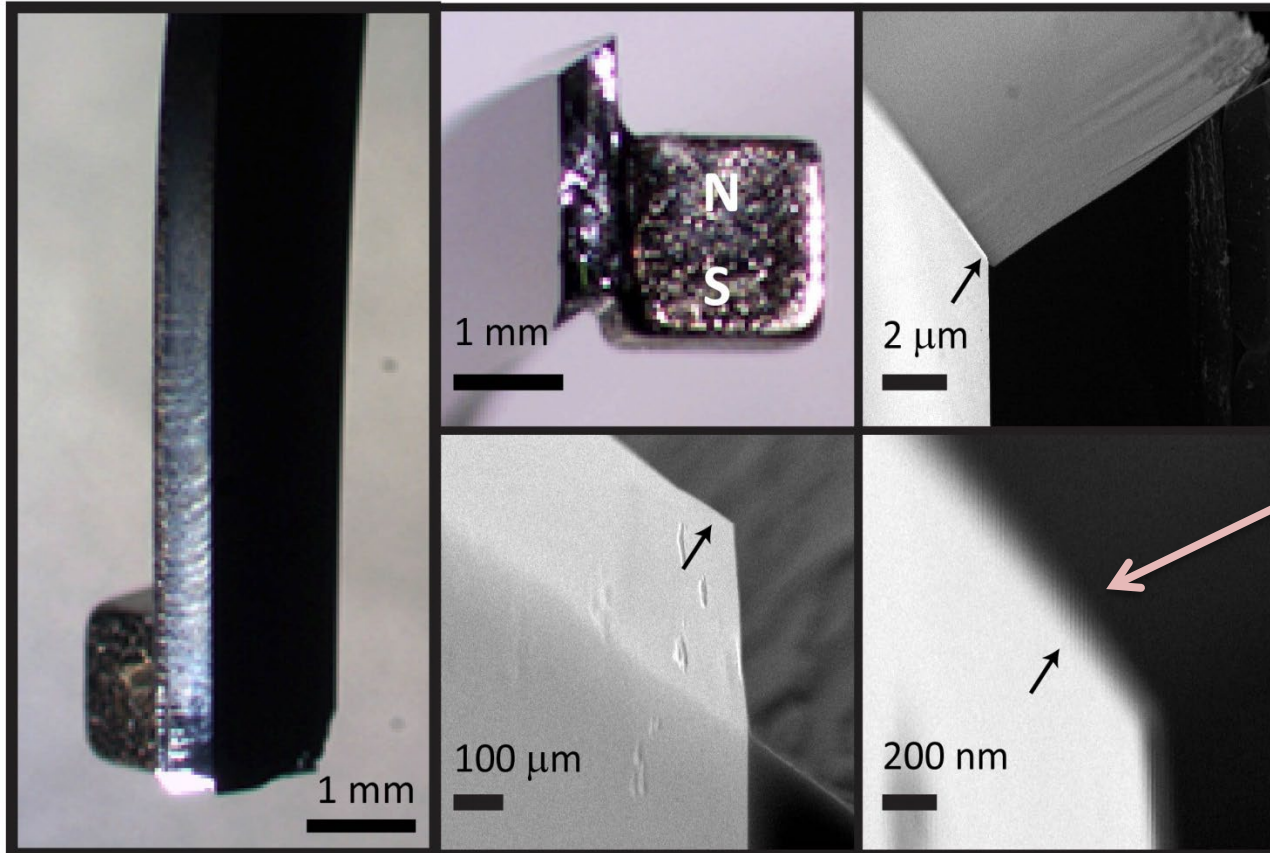
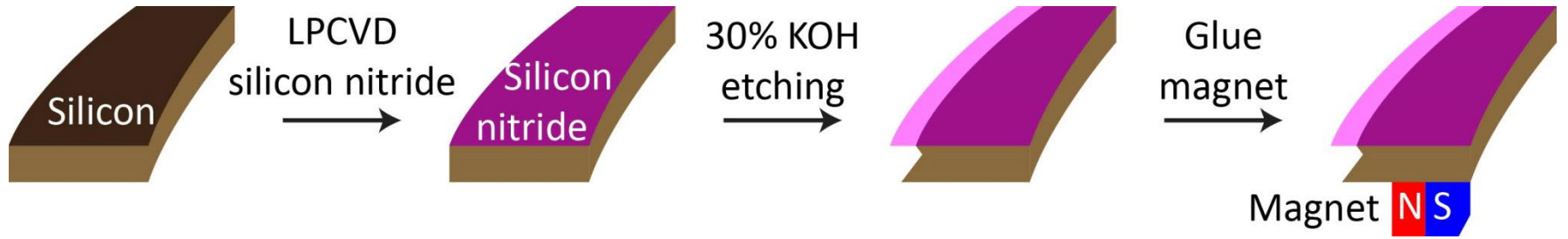


Rotating force



Varying force

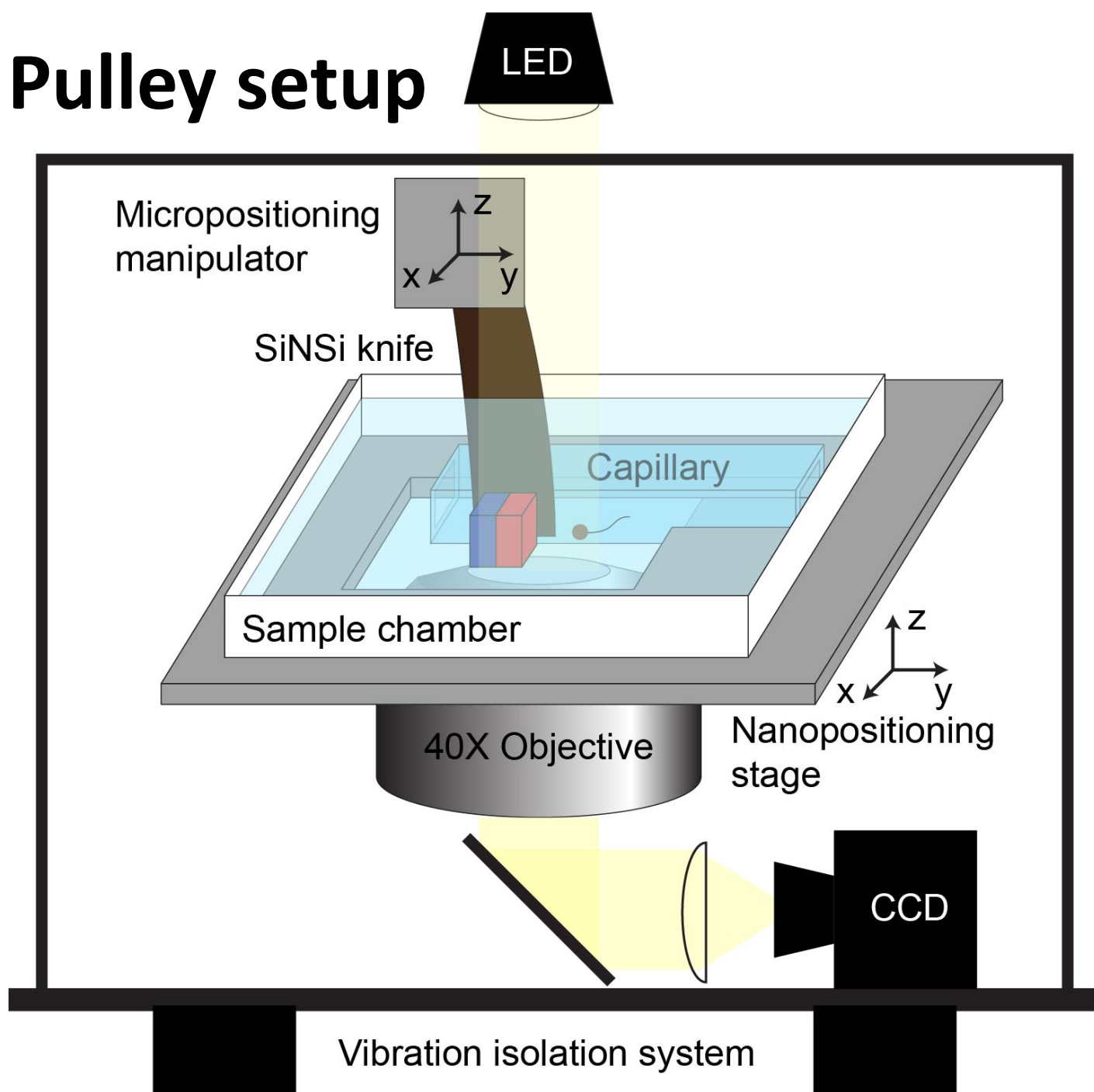
Silicon nitride knife



*Stereomicroscope
and SEM Images*

*Nanoscopically
smooth edge*

DNA Pulley setup



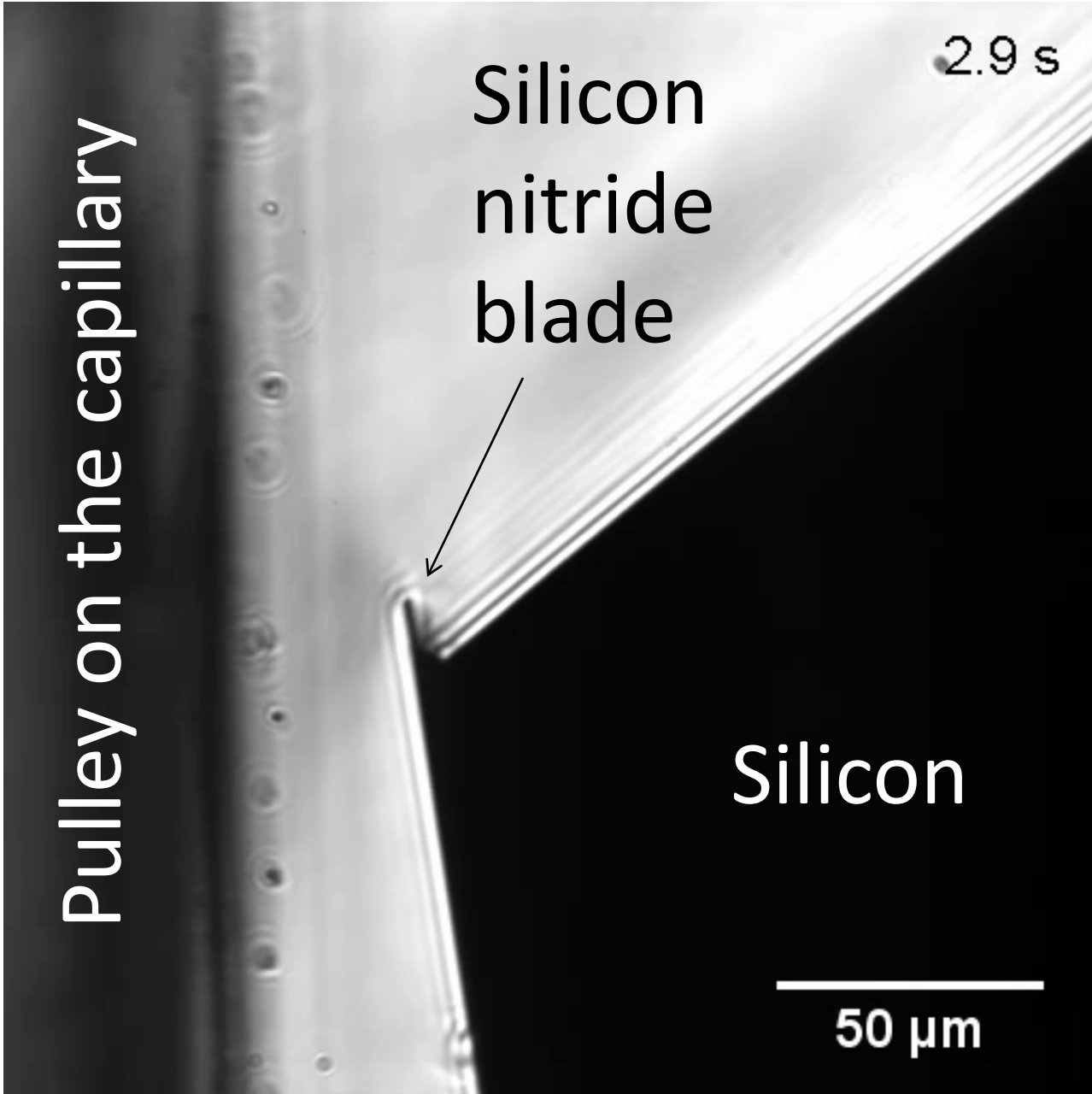
Pulley on the capillary

Silicon
nitride
blade

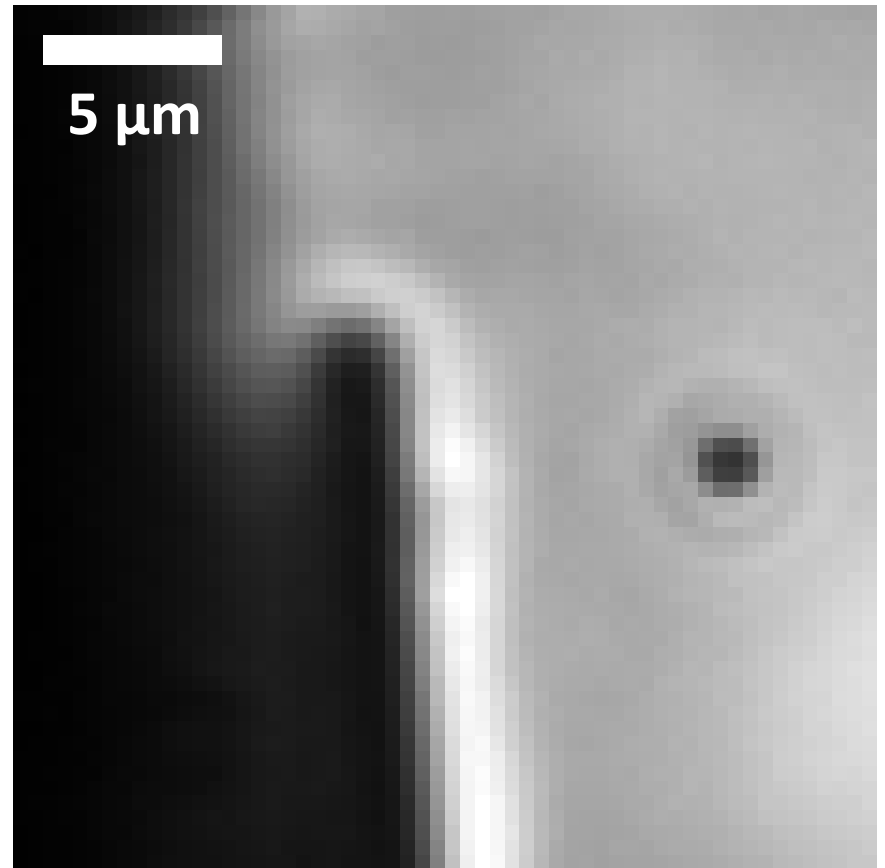
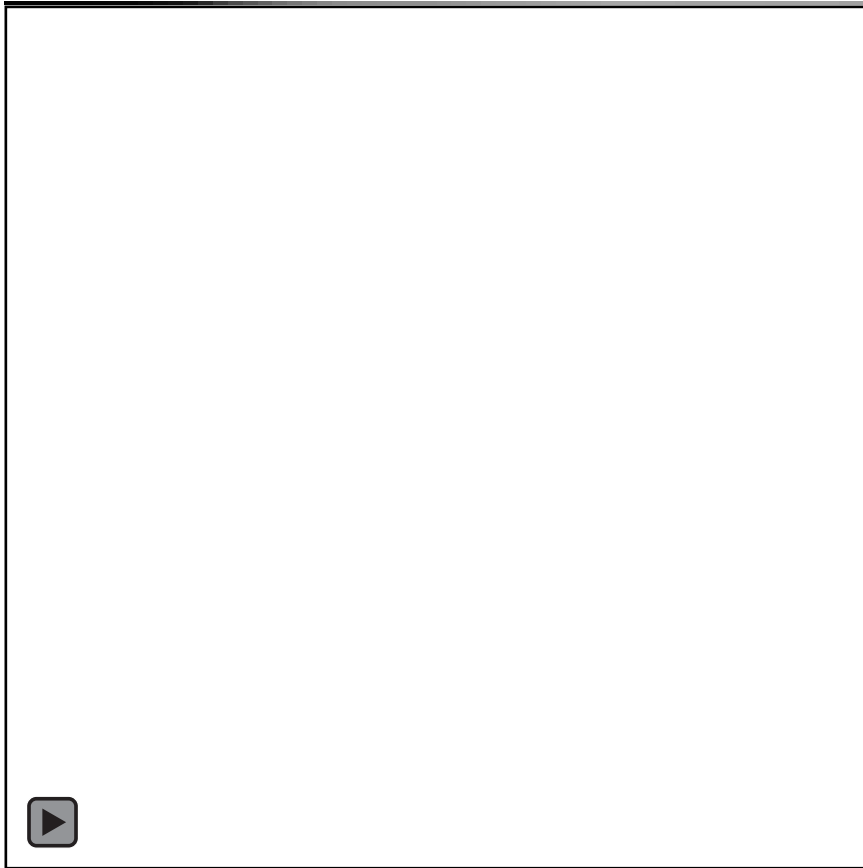
2.9 s

Silicon

50 μm

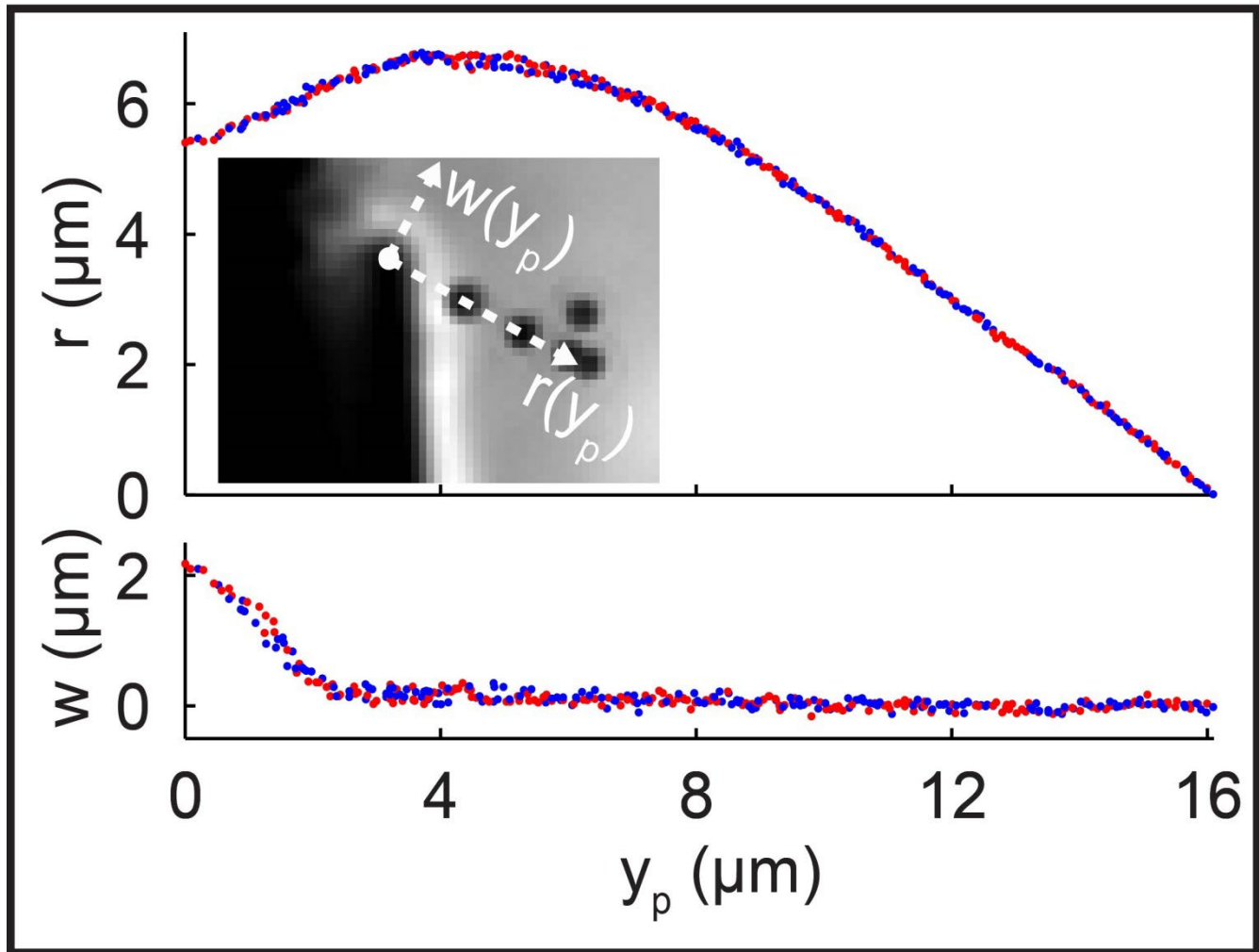
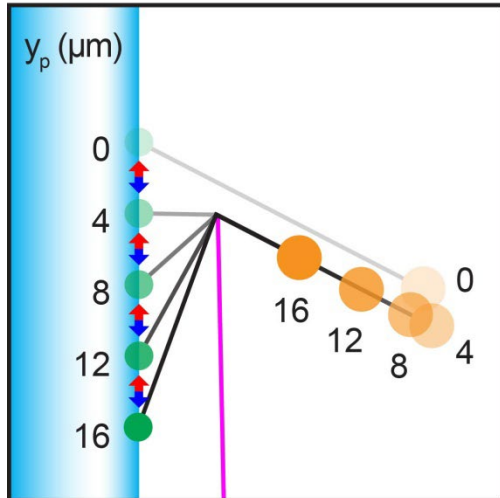
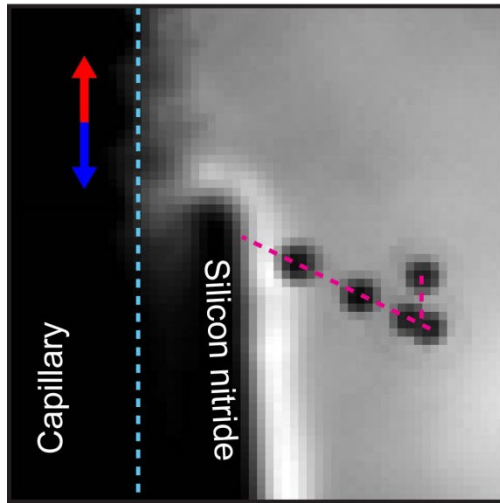


Simple scanning

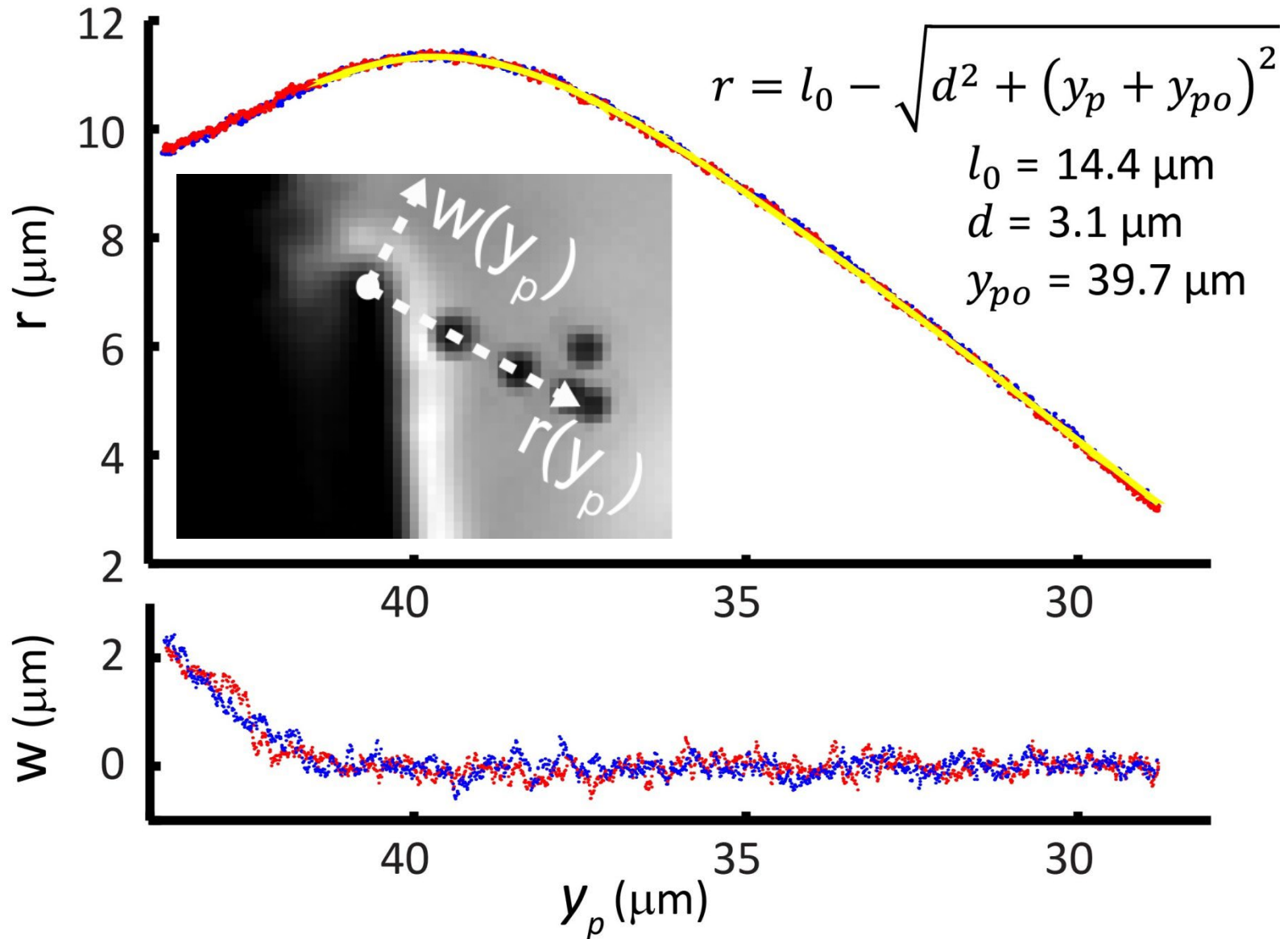


$$r = l_o - \sqrt{d^2 + (y_p + y_o)^2}$$

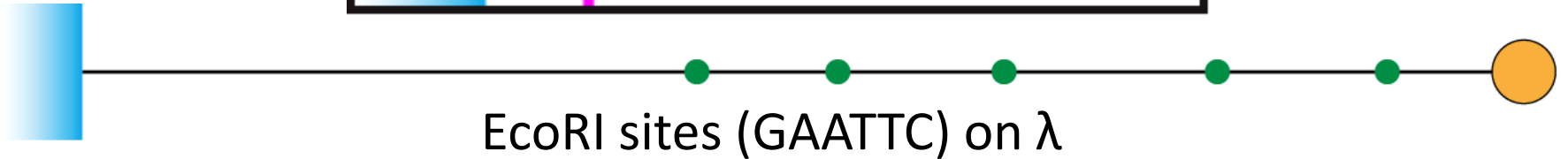
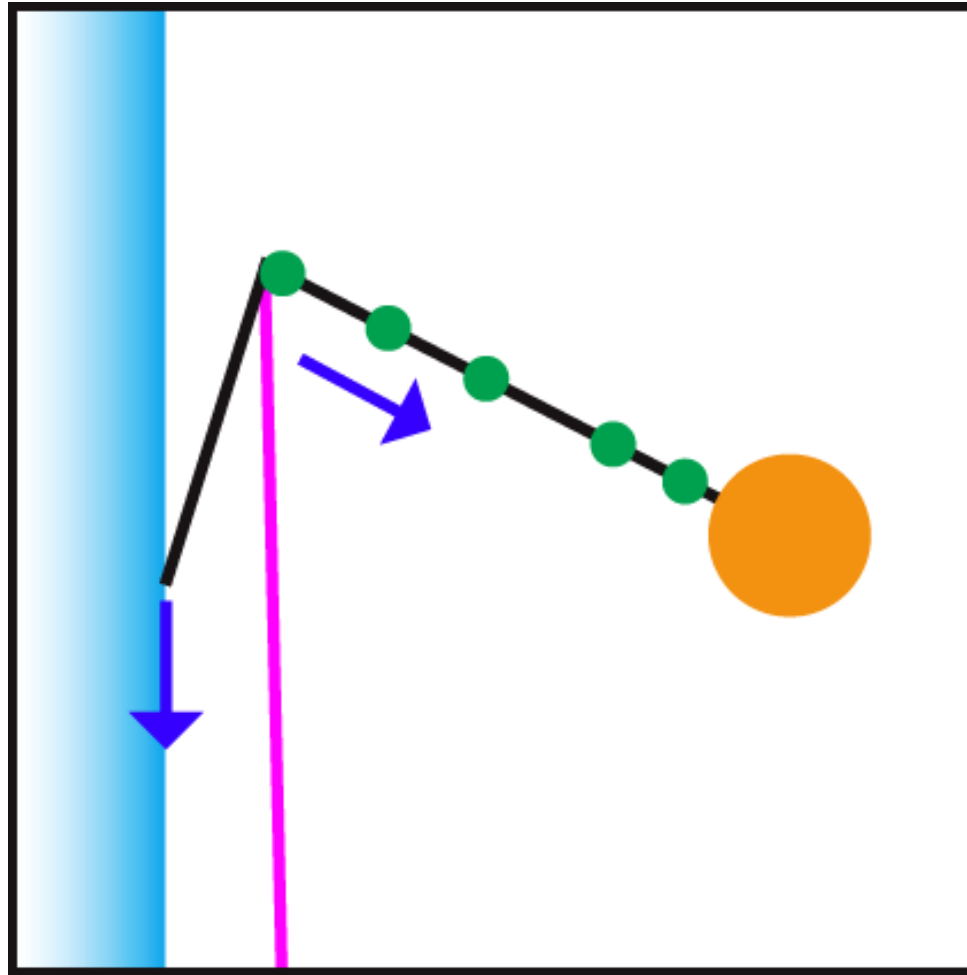
l_o : total extension; d : capillary distance; y_o : offset in y



Bead trajectory

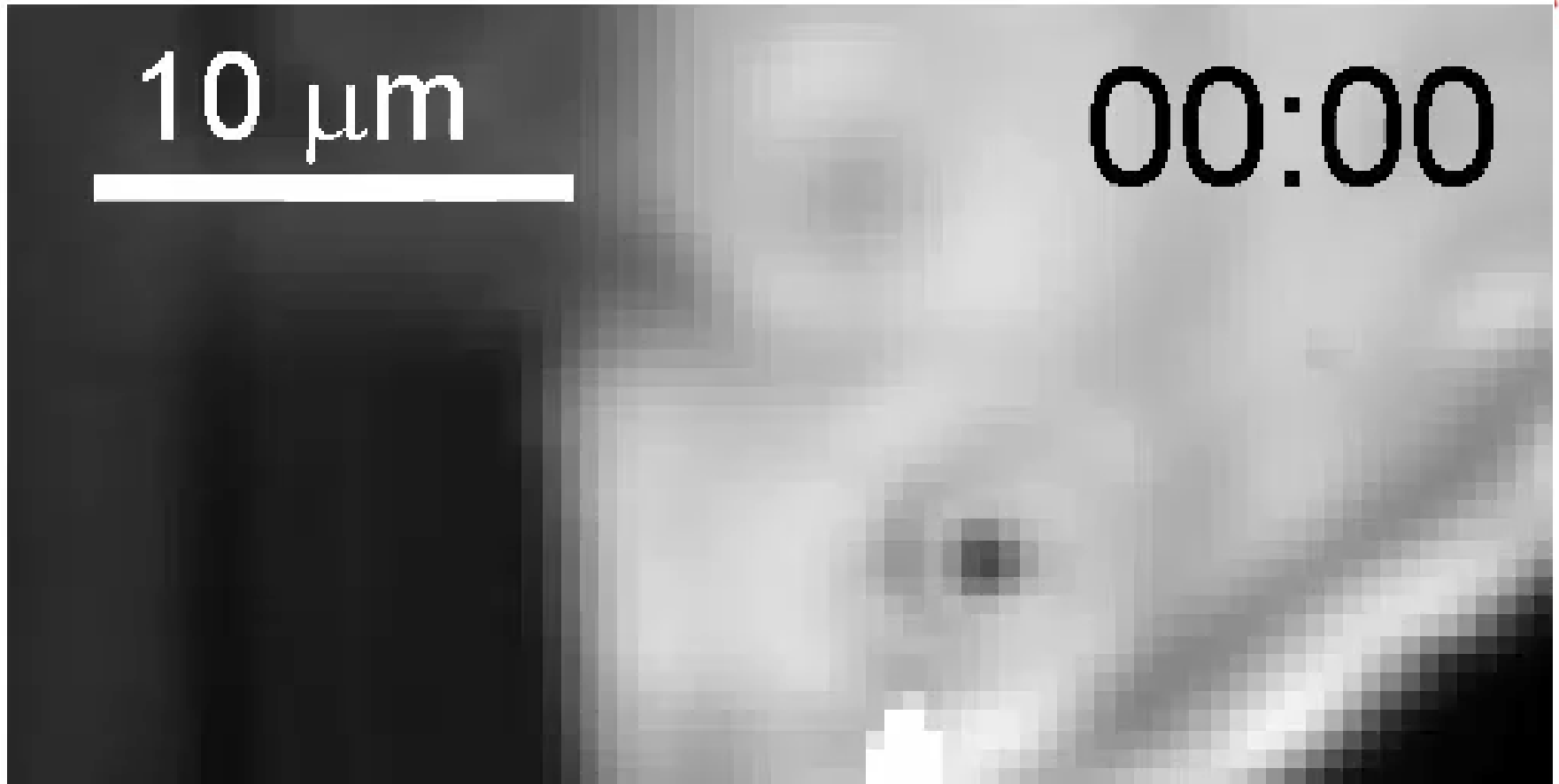


Fiducial marker: EcoRI



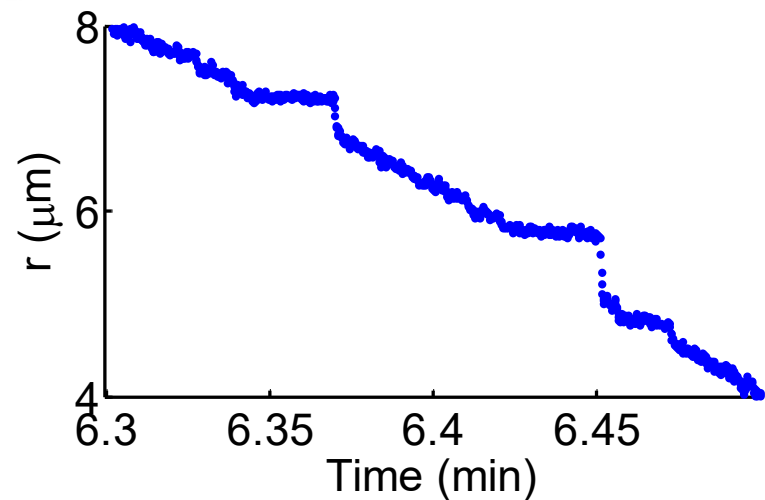
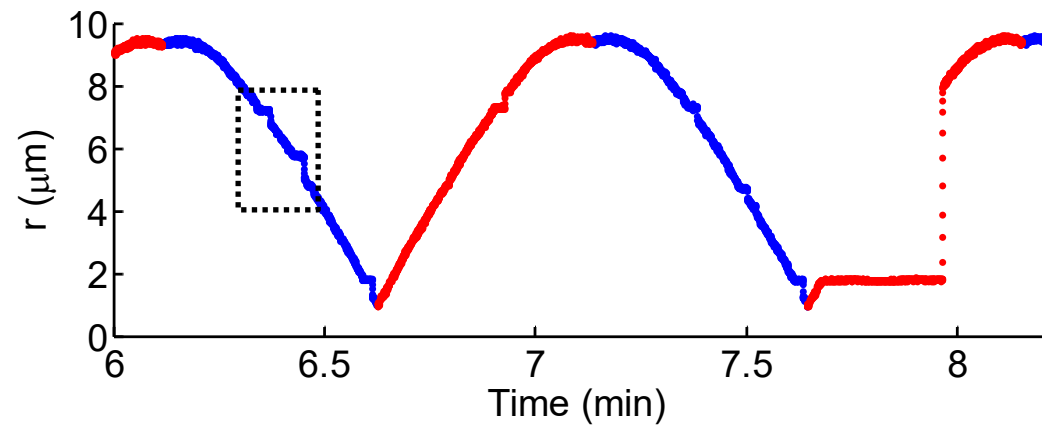
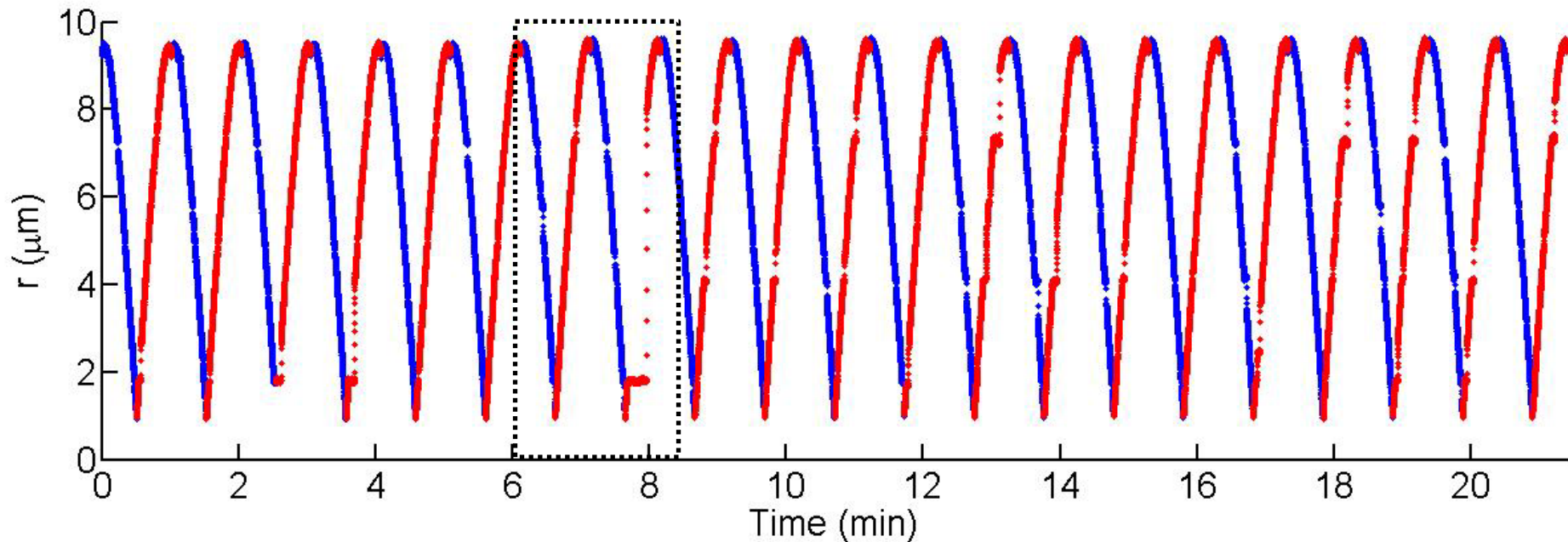
EcoRI sites (GAATTC) on λ

EcoRI-incubated λ DNA

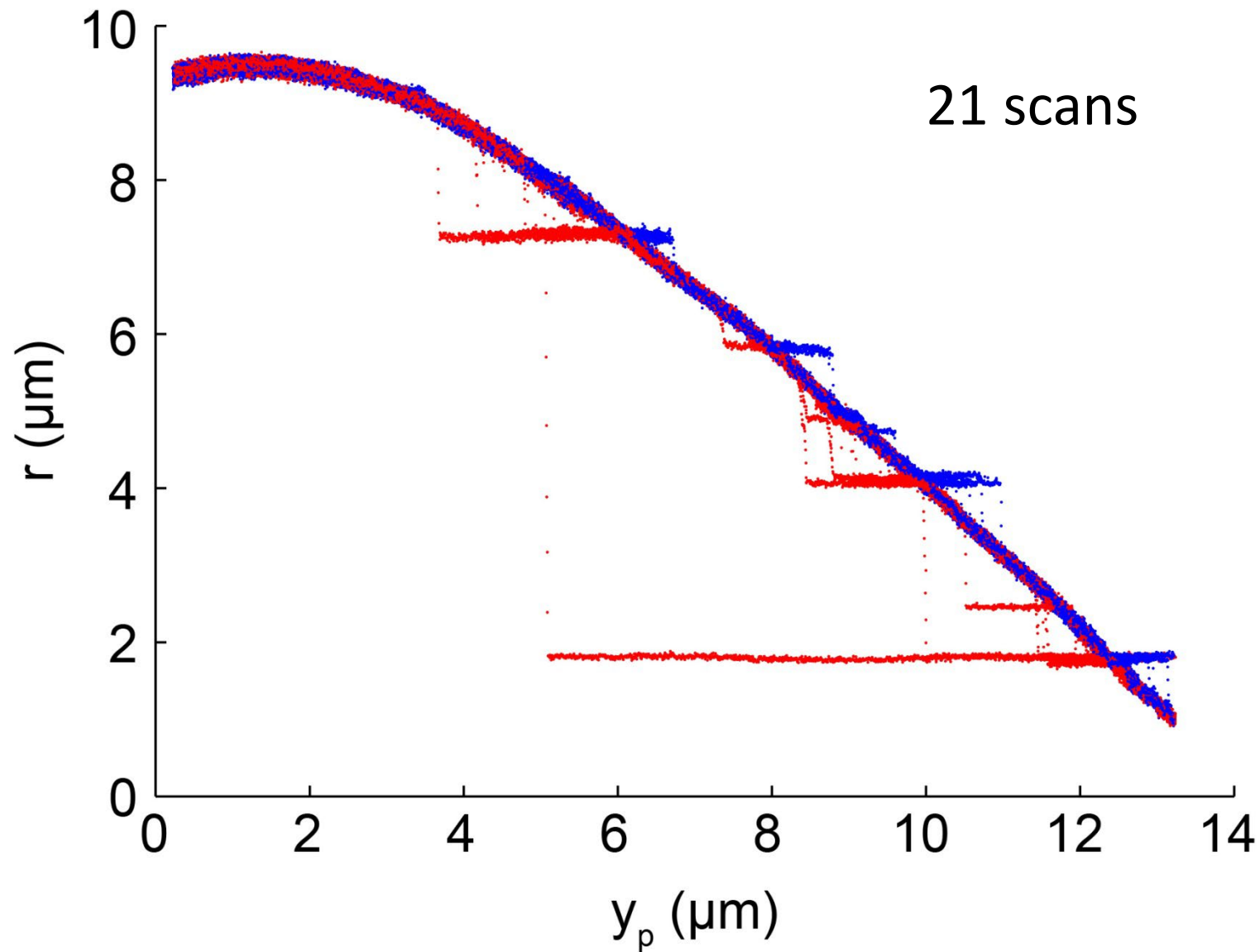


λ DNA with 10 nM EcoRI, 1 mM CaCl_2

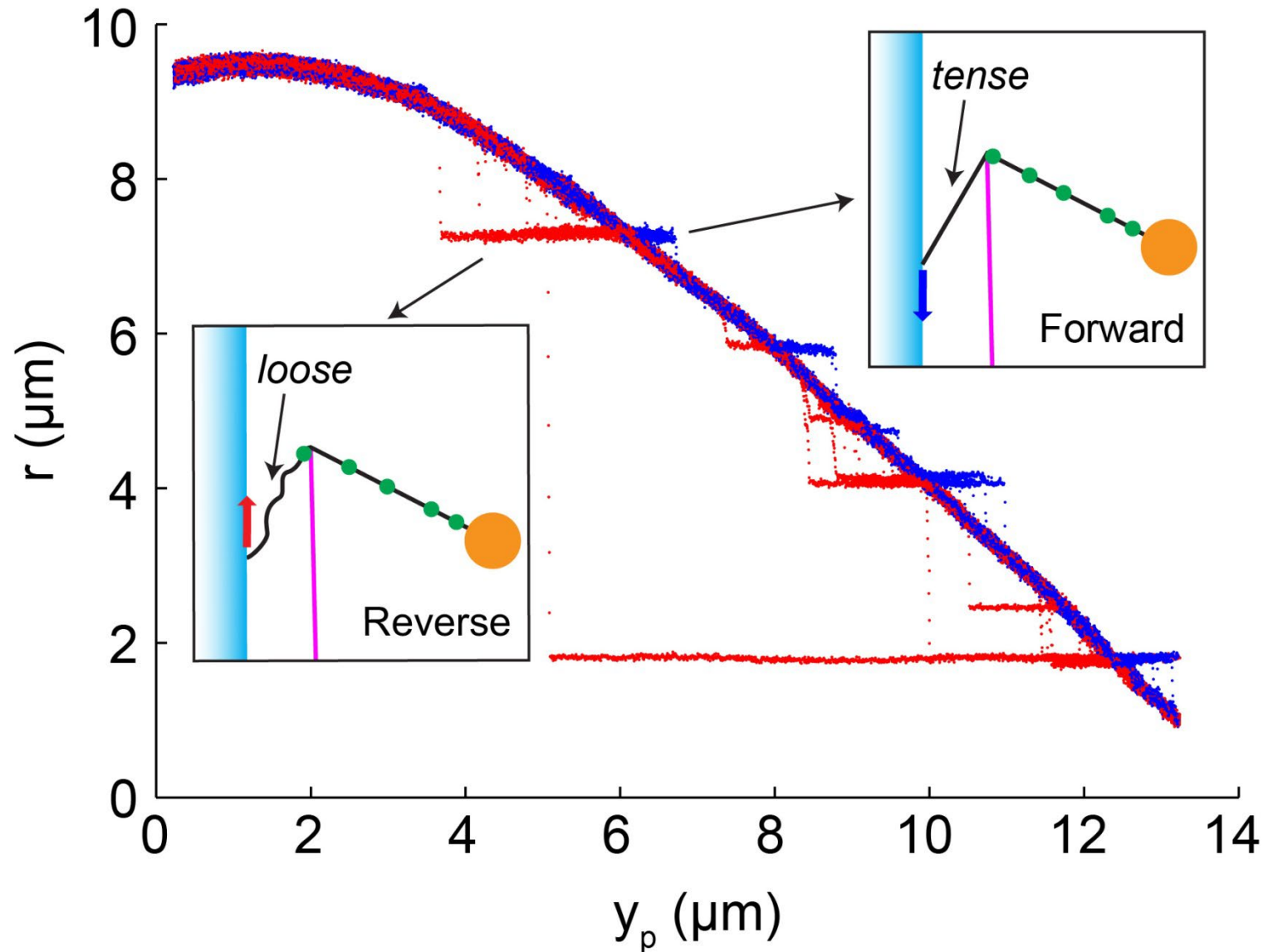
EcoRI-incubated λ DNA



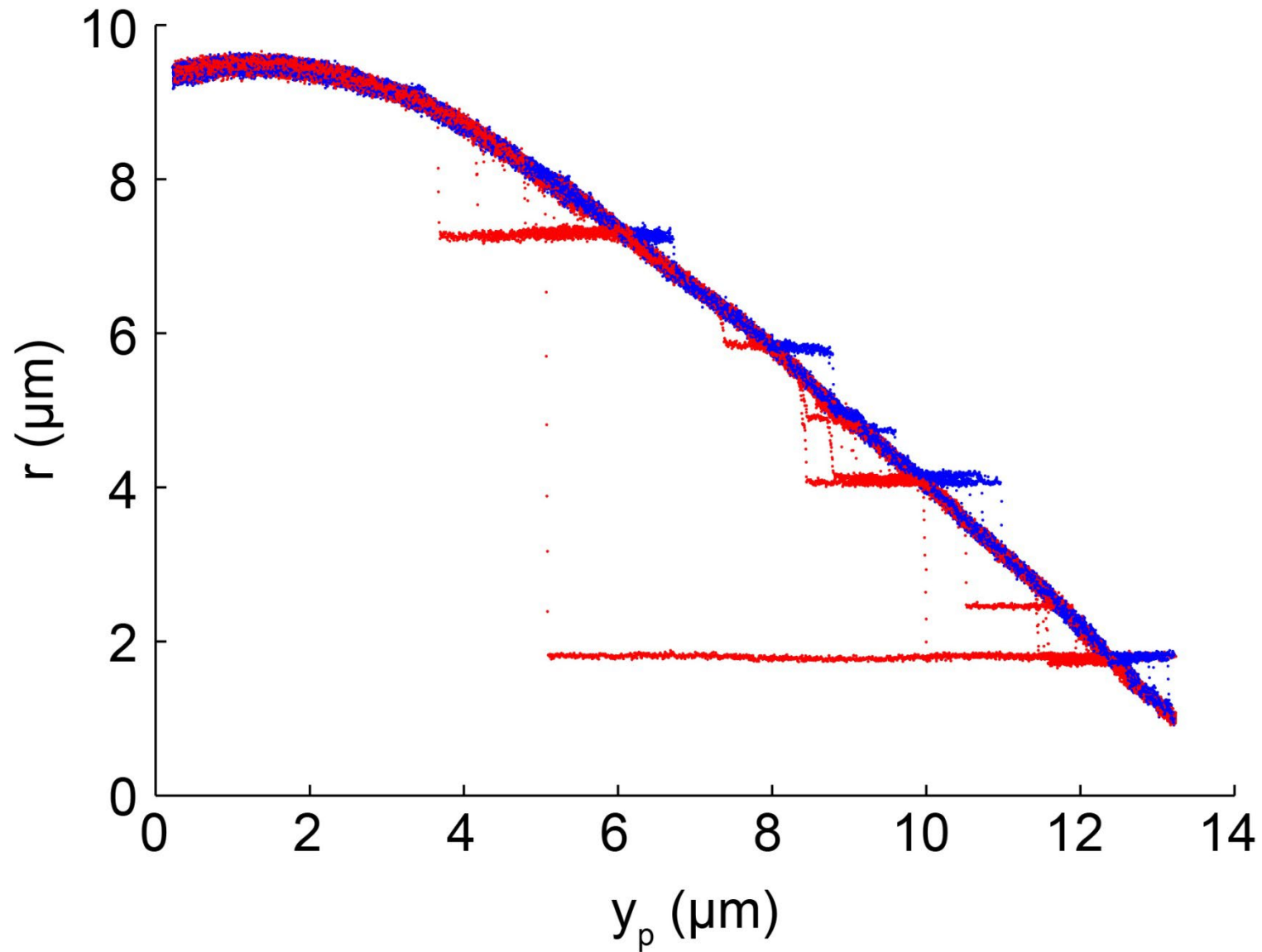
EcoRI on DNA



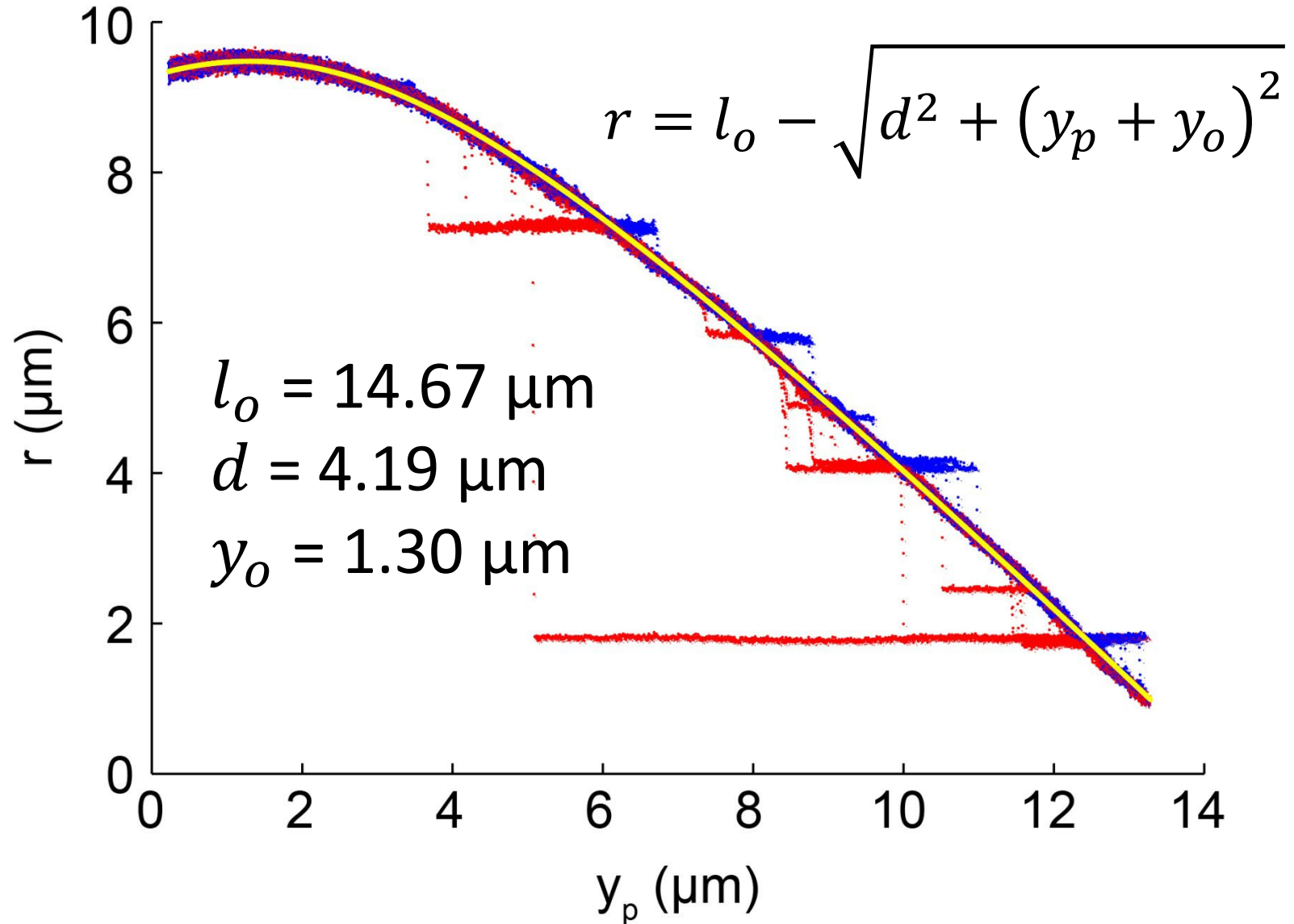
Asymmetry in the direction



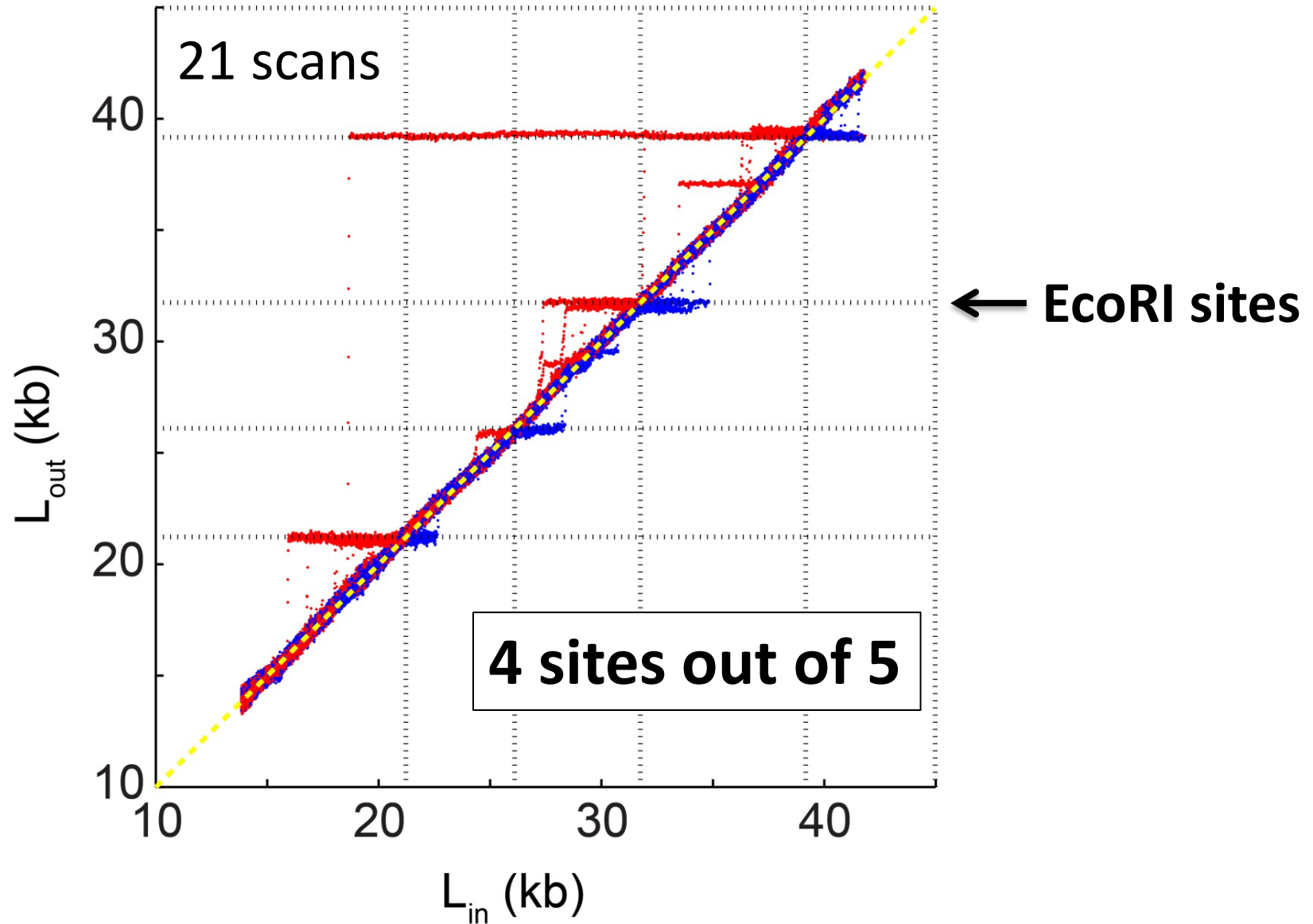
EcoRI on DNA



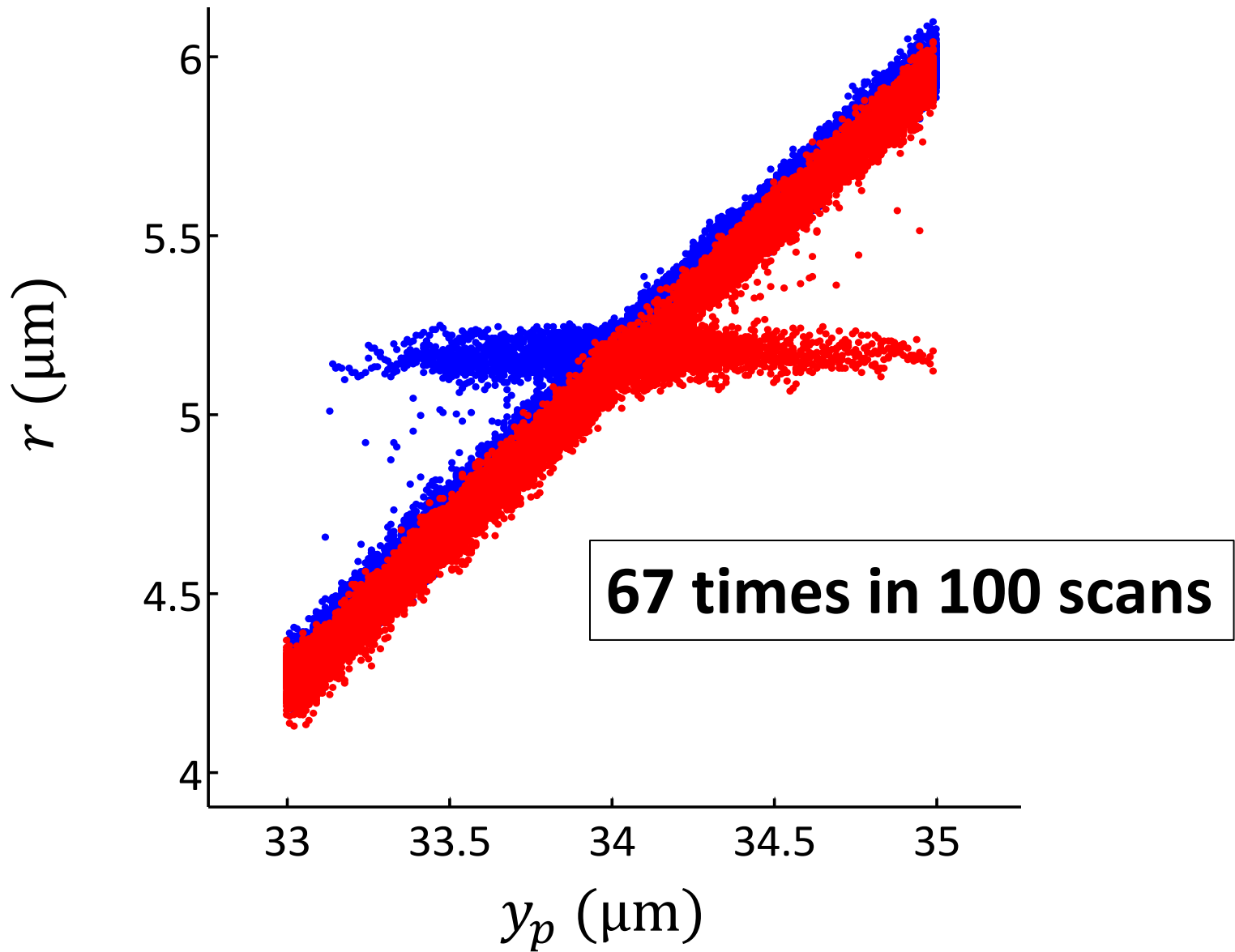
EcoRI on DNA



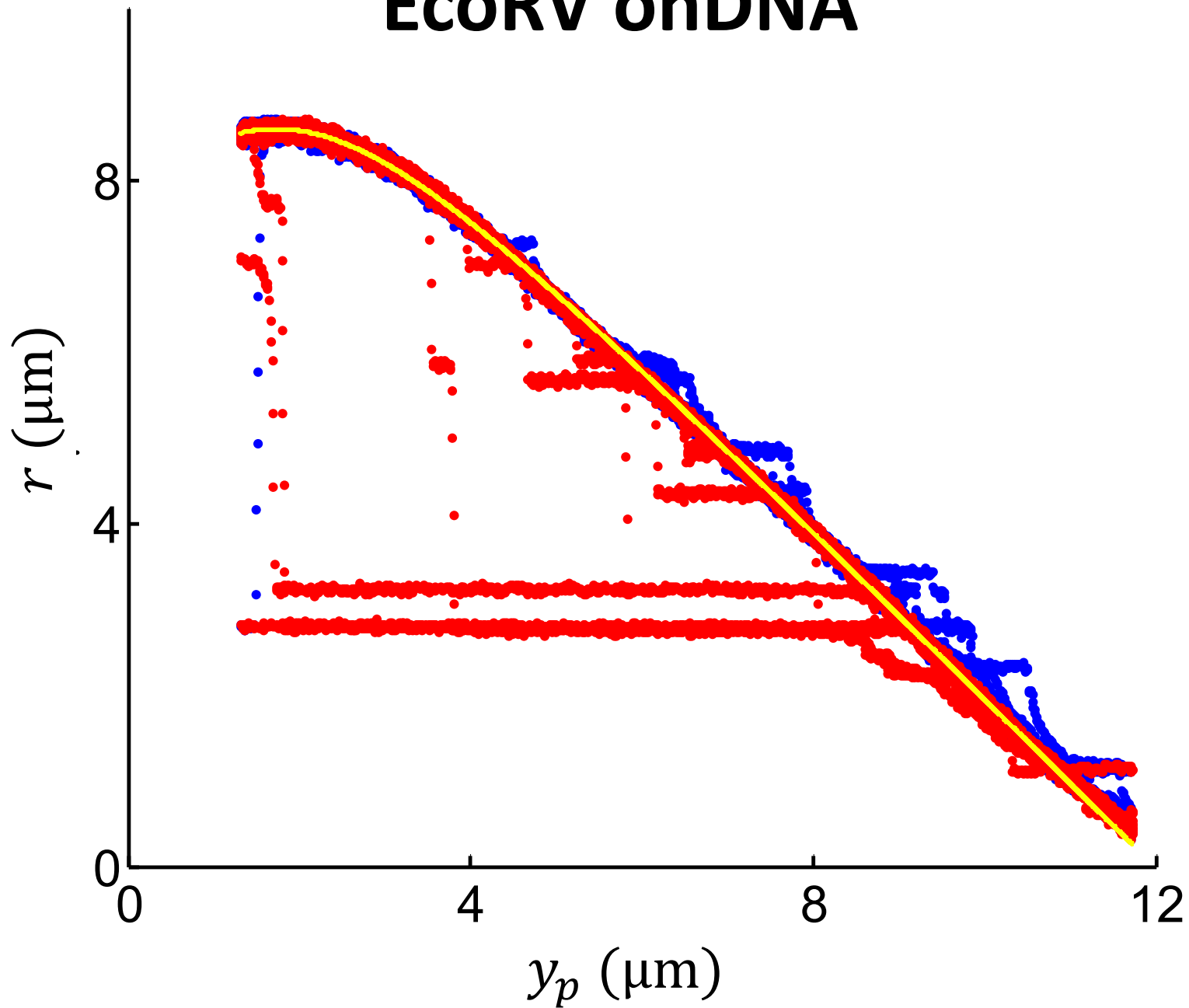
Mapping recognition sites



Detection efficiency

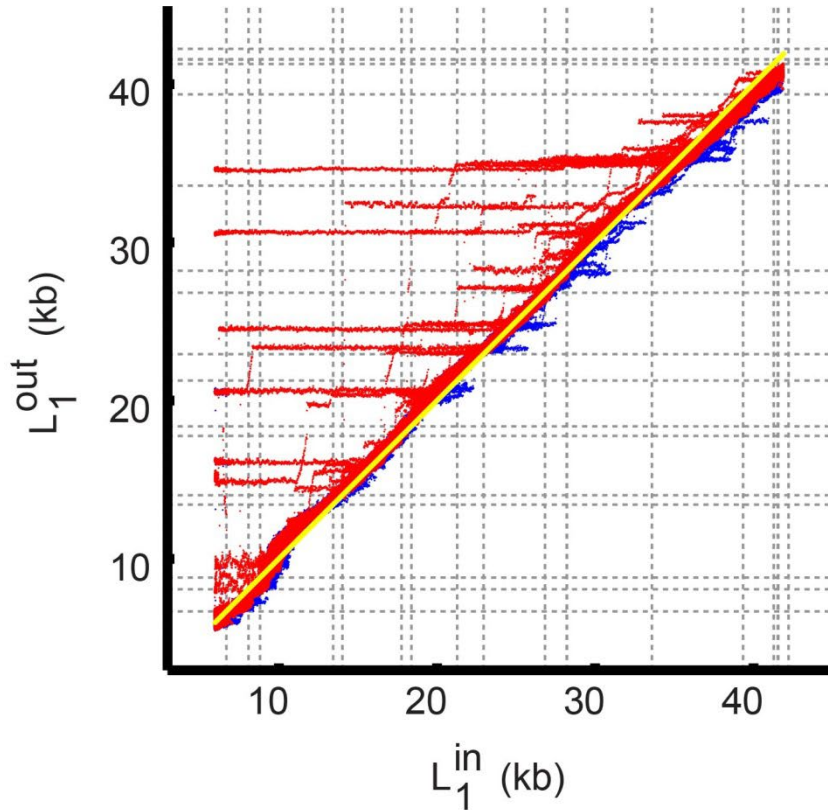


EcoRV onDNA



EcoRV bound to DNA

Pulley #1



Pulley #2

